GCA2020 – Global Conference on

² Aquaculture

- 3 Aquaculture for Food and Sustainable Development
- 4 23-24 September 2021

Key messages

- Research to reduce global warming from aquaculture should be a priority.
- The aquaculture sector should continue to improve and promote sustainable and environmentally sound practices.
- The ocean should be used more effectively, efficiently and intelligently to produce food for a growing human population.
- In areas with a high potential for shellfish or seaweed production, aquaculture needs to be stimulated by improving licensing procedures, educating the public, and implementing smart technologies.
- Aquaculture development in Africa and Latin America should be actively stimulated, with the aim of distributing aquaculture more evenly across countries on each continent and between continents globally.
- The use of technologies to collect, treat, and re-use aquaculture wastes in a responsible way should become the standard in any aquaculture operation throughout the industry.
- The aquaculture industry should aim to integrate aquaculture in nutrition-sensitive food systems, and actively advertise the contribution of aquaculture products to a nutritious diet.
- The aquaculture industry should explore the use of smart technology to improve farming efficiency and reduce production costs across the industry, especially for shellfish and seaweed culture.
- 5 6

Aquaculture systems

7 Abstract

8 Since 2000, the aquaculture industry has become well integrated in the global food system. Aquaculture

- 9 production has tripled, reaching a production of 85 million metric tonnes (Mt) of aquatic animals and 35
- 10 Mt of aquatic plants in 2019. Aquaculture systems are highly diverse, producing globally equal live
- weight amounts of fed species and extractive species. In Asia and Africa, inland aquaculture provides thebulk of aquaculture production, while in the Americas, Europe and Oceania, marine aquaculture
- 13 dominates. The realized growth is mainly due to intensification, use of more and better feeds, improved
- 14 production management and increased attention to biosecurity.
- 15
- 16
- 17

18 Challenges to the industry include reducing the impact from fed aquaculture on pollution and global

19 warming. Both fed and extractive aquaculture need to pay more attention to scaling, site selection and

- 20 the health of the wider production environment. In terms of land use, aquaculture is more efficient than
- 21 terrestrial animal production, but water use remains a challenge, and more attention should be given to
- 22 water recycling in land-based systems, reducing water consumption and facilitating nutrient recovery and
- 23 re-use.

24 Future development should focus on making the aquaculture industry climate neutral and reduce

25 environmental impacts, both inland and at sea. More attention must be given to making aquaculture an

26 important part of the food system on all continents, like is the case in Asia today. Increasing the global

27 production volume should not be a major goal by itself. Integration of aquaculture into local nutrition-

sensitive, circular and sustainable food systems should become the major driver for future aquaculturesystem development.

30 1 Introduction

31 This paper summarizes the development of aquaculture between 2000 until today. Aquaculture systems 32 are highly diverse with presently ca. 425 species in production (Naylor et al., 2021). As each one of 33 these species can be cultured in different production systems ranging from extensive to super-intensive, 34 the number of possible systems is enormous. Therefore, we define an 'aquaculture production system' in 35 broad terms as a production environment in which interventions aim to enhance the value and/or the 36 amount of biomass produced, to the benefit of the people and communities organizing and executing 37 these actions. Actions range from stimulating recruitment of desired species, to stocking fast growing 38 strains, protecting against disease, fertilization, feeding and creating production environments like ponds, rafts supporting hanging or floating ropes or long-lines, cages, pens, raceways and recirculating 39 40 aquaculture systems that allow control on resource use efficiency to a higher degree than in natural 41 systems.

42 Aquaculture can only optimize its contribution to society and nature through responsible use of resources 43 and strong integration within the global food system. Claims by aquaculture for freshwater and for space 44 on land and at sea should be coordinated with claims from agriculture, urbanization, industry and nature 45 at regional or water basin scales. These are complex questions that require multi-objective analysis 46 (Pelletier et al., 2018). Similar complex dilemmas exist for the selection of ingredients for fish feeds to 47 replace animal-based resources with plant-based ones, finding a right balance with impacts on the 48 environment, biodiversity, climate change, competition with human foods, fish health, fish welfare and 49 nutritional quality of aquaculture products.

50 FAO holds every 10 years a Global Conference on Aquaculture. The next meeting will be held September 51 2021, following up on meetings organized in Thailand in Phuket in 2010 (Subasinghe et al., 2013) and 52 Bangkok in 2000 (NACA/FAO, 2000). The goal of these meetings is to set priorities for aquaculture 53 development during the next decade. This paper summarizes the present state of aquaculture looking 54 from a systems perspective, and set priorities for the future. Because the Phuket declaration in 2010 was 55 an extension of the Bangkok declaration in 2000, providing a small extension to the recommendations 56 given 10 years earlier during the Bangkok conference, we analyze progress since 2000 with focus on the 57 last decade.

- 58
- 59
- 60
- 61
- 62
- 63

- 64 The manuscript is organized in 3 major sections:
- 65 <u>1</u> Aquaculture development since 2000
- Addresses how well the sector developed during the last decades. Considering the large differences
 that exist in system requirements for the production of fish, crustaceans, molluscs, and seaweed,
 these are reviewed separately.
- 69 <u>2</u> Current issues and challenges in aquaculture
- Addresses what are the current issues and challenges facing the industry today? Harmful and
 positive impacts of aquaculture production systems on the environment, society, and the ability to
 produce within planetary boundaries are reviewed.
- 73 <u>3</u> Key developments for the next decade
- 74 Discusses which key developments in the aquaculture sector need prioritization to deliver sustainable75 growth during the next decade?

3

76

77 2 Aquaculture development since 2000

78 2.1 Development of aquaculture production (2000 – 2019)

Aquaculture is one of the fasted growing food sectors worldwide. During the last 20-30 years, the sector 79 80 showed a constant and significant increase in the contribution of commercial and industrial aquaculture to 81 global production. Also small-scale and medium-scale aquaculture enterprises benefited from this growth 82 of the aquaculture industry. By growing and commercializing, the sector increased its contribution to food 83 security, income, and trade. Growth has been mainly in Asia, with other continents lagging behind in 84 realizing the potential of aquaculture to contribute to food production and food security. There are, 85 however, signs of change. During the last decade, growth rates in production were higher in Africa and 86 Latin America than in Asia, showing a growing interest in food production through aquaculture outside 87 Asia. Naylor et al. (2021), looking at all-sub-sectors of the aquaculture sector, state that during the last 88 decades aquaculture became better integrated within the global food system, while realizing large 89 improvements in environmental performance. Responsible aquaculture today is considered a legitimate 90 user of resources that improves livelihoods and contributes to environmental enhancement.

91 Between 2000 and 2019, world aquaculture production increased from 43.0 to 120.1 million metric ton 92 (Mt), an increase close to 180% (Table 1). In 2019, animal production through aquaculture reached 85.4 93 Mt and the production of aquatic plants 34.7 Mt. Asia, with China as the main producer, has always been 94 the continent leading aquaculture production. Asia was responsible for 90% of global production in 2000 95 and increased its share to 92% in 2019. The other continents are lagging behind, with the Americas, Europe, Africa and Oceania producing respectively 3.5, 2.7, 2.0 and 0.2 % of the global production in 96 2019. The large difference in production between Asia and the other continents indicates there is a vast 97 98 potential for aquaculture development outside Asia.

The contribution of Europe and Oceania to global aquaculture production is declining, is more or less steady in the Americas, and has doubled in Africa between 2000 and 2019. In the Americas, the average annual growth rate (AAGR) of aquaculture between 2000 and 2010 was 3%, but increased to 6% between 2010 and 2019, mainly due to growth in Central and South America. Over the same period, the AAGR in Africa was also 6% (Table 1, Figure 1; (FAO/FishStatJ, 2021; Garlock et al., 2020). With higher growth rates in Africa and the Americas than in Europe, these continents most likely will in the near future overtake Europe as the second largest aquaculture producer.

- 106
- 107
- 108
- 109

110 If interest in aquaculture spreads to more countries than is presently the case in Africa and Latin America

(Figure 1), and the annual growth rate remains above average as is presently the case, then these

112 continents will increase their contribution to global aquaculture production during the next decade.



Figure 1

Growth in aquaculture production quantity (%) between 2006 and 2016 for countries with production greater than 100 000 MT. Reproduced from <u>Garlock et al., 2020</u>.

116 Table 1:

117

Global aquaculture production in 2000, 2010, and 2019 by continent (<u>FAO/FishStatJ, 2021</u>). AAGR: average annual growth rate calculated for the period indicated.

	2000		2010		% AAGP	2019		% AAGR
	production	% global	production	% global	2000 - 2010	production	% global	2010 - 2019
Country (Name)	(Mt)	production	(Mt)	production	2000 2010	(Mt)	production	2010 2013
Asia	38.9	90	71.3	91.4	6.2	110.0	91.6	4.9
Americas	1.5	3.4	2.5	3.2	5.7	4.2	3.5	5.8
Europe	2.1	4.8	2.5	3.2	2.1	3.2	2.7	2.8
Africa	0.5	1.0	1.4	1.8	12.2	2.4	2.0	5.9
Oceania	0.1	0.3	0.2	0.3	3.9	0.2	0.2	1.1
World	43.0	100	78.0	100	6.1	120.1	100	4.9

118

Between 2000 and 2019, the contributions of finfish, crustaceans, and other aquatic animals to global aquatic animal production through aquaculture increased 2, 7, and 0.5%, respectively. In contrast, the share of molluscan culture to global aquatic animal production declined 9% during the same period. Overall, the contribution of aquatic plants to the global aquaculture production increased by 4% between 2000 and 2019, concurring with a decline from 75 to 71% of the contribution of aquatic animals to global aquaculture production (Table 2).

125 Most striking is the 5.2-fold increase in global production of crustaceans between 2000 and 2019. This is 126 mainly due to their high market value. In 2000, the average price on a global scale of finfish was 1.2 127 US\$/kg compared to 4.8 US\$/kg for crustaceans, a 4-fold price difference. By 2019, the price of finfish 128 and crustaceans increased to 2.6 and 7.3 US\$/kg, respectively, representing nearly 3-fold price premium 129 for crustaceans compared to finfish (<u>FAO/FishStatJ, 2021</u>). In spite of the much higher price for crustaceans 130 compared to finfish, the production cost of crustaceans is substantially higher, with higher system 131 requirements and input costs. When all goes right, crustacean farmers earn on average more than finfish 132 farmers, but commercial risks are also higher given the increased potential for culture failure due to disease 133 or environmental disasters. Nevertheless, considering the present high demand for crustaceans in Asian markets, further fast growth of crustacean production is expected during the next decade with minor 134 135 impact on the price premium for crustaceans.

136 In contrast to the global finfish production which nearly tripled between 2000 and 2019, global production 137 of molluscs only doubled. Also the average price for molluscs doubled from 0.9 US\$/kg in 2000 to 1.8 138 US\$/kg in 2019. On average, the price of molluscs is 2/3 the price of finfish, a ratio that remained similar 139 between 2000 and 2019 (<u>FAO/FishStatJ, 2021</u>). The low price, in combination with a low consumer 140 demand, might explain in part the slow development of molluscan aquaculture, although there is 141 considerable room for further development.

142

144 The vast majority of finfish, except for ca. 8 Mt of carp species, and crustaceans are fed formulated pelleted

145 feed, while molluscs are not fed (FAO, 2020). Seaweeds are grown at sea, nearly always without fertilizer 146 addition, although in coastal waters or bays, there might be substantial nutrient runoff from land (Mahmood

147 et al., 2016b).

- 148 Therefore, we refer to finfish, except 8 Mt of carps, and crustaceans as fed species and to 8 Mt carps, 149 molluscs, and seaweed as extractive species. The main species group 'other aquatic animals' are also
- 150 considered fed species (Table 2).

151

152

153

Table 2: Global aquaculture production in 2000, 2010, and 2019 by main species group (FAQ/FishStatJ, 2021) (Mt, million metric ton).

	20	2000 2010			20	% AAGR		
Main species grouping	Production (Mt)	% global production	Production (Mt)	% global production	2000 - 2010	Production (Mt)	% global production	2010 - 2019
Finfish ¹	20.8	64	37.7	65	6.1	56.3	66	4.5
Crustacea ¹	1.7	5	5.5	9	12.5	10.5	12	7.5
Molluscs ¹	9.8	30	13.8	24	3.5	17.6	21	2.7
Other aquatic animals ^{1, 3}	0.2	0	0.8	1	17.7	1.0	1	2.4
All aquatic animals ²	32.4	75	57.8	74	6.0	85.4	71	4.4
All aquatic algae ²	10.6	25	20.2	26	6.7	34.7	29	6.2
Fed species ⁴	19.7	46	38.7	50		59.8	50	
Extractive species ⁵	23.3	54	39.3	50		60.3	50	
All Species	43.0	100	78.0	100	6.1	120.1	100	4.9

154

¹ Percentage (%) global production calculated against 'All aquatic animals' production; ² % global production 155

calculated against 'All species ' production'; ³ Other aquatic animals include amphibians, reptilians, and aquatic 156 157 invertebrates; ⁴ Fed species include finfish minus non-fed carps, Crustacea, and Other aquatic animals; ⁵ 158

Extractive species include non-fed carps, Molluscs, and All aquatic algae.

159

160 2.1.1 Fed species

161 2.1.1.1 Production overview

Global finfish production through aquaculture increased from 20.8 million Mt in 2000 to 56.3 Mt in 2019, 162 163 an increase of 170%. Global crustacean production increased from 1.7 to 10.5 Mt during the same period, 164 an increase of 520%. Combined, the share of crustaceans and finfish to global animal production through 165 aquaculture grew from 69% in 2000 to 78% in 2019; however, including seaweed, the fed 166 species:extractive species ratio of aquaculture by volume is 50:50. Since 2000, when this ratio was 46:54 167 the importance of fed species gradually increased, a trend that will continue in the near future (Table 2). 168 When considering only animal production through aquaculture, then the fed species:extractive species 169 ratio of aquaculture was 70:30 in 2019, whereas this ratio was 61:39 in 2000.

170 In total, 56 Mt of finfish was produced in 2019, representing 66% of the global animal production through 171 aquaculture, of which 86% is produced inland and 14% in marine areas (Table 3). The importance of inland 172 aquaculture is mainly due the dominant position of Asia worldwide, who produces 91% of its aquatic animal 173 production inland. However, with the exception of Africa where 87% of aquaculture production is inland, 174 in other continents marine aquaculture is more important. The Americas, Europe and Oceania culture respectively 50, 79 and 95% of their finfish in marine areas. Inland aquaculture consists mainly of finfish 175 176 culture representing on all continents \geq 90% of global inland production of aquatic animals, reaching 100% 177 in Africa and Europe. Looking at marine aquaculture, the picture is much more diverse. Asia for instance 178 produces more crustaceans than finfish in marine aquaculture (20 vs. 17%). On other continents, the 179 contribution of finfish to marine aquaculture production varies between 39 and 96% (Table 3).

Globally, 60% of crustaceans are produced in marine areas, with large variations between continents. In total, 10.5 Mt was produced in 2019, representing 12% of the global production of aquatic animals through aquaculture. Of the inland aquaculture production in the Americas, 8% are crustaceans, while for marine aquaculture this is 36%. In Asia, the contribution of crustaceans to marine aquaculture is 20%, similar to the global average. On the other continents, the productions of crustaceans are negligible to very small (Table 3).

Shrimp production is mainly destined for export in many producing countries while with the exception of high value finfish species (e.g. salmon, grouper), finfish are consumed domestically or in the region of production. For crustaceans, the international trade volume and traffic varies between the regions of production, depending on shifts in the balance between supply and demand.

190 Table 3:

191

Inland and marine finfish and crustacean production (thousand metric ton) through aquaculture by continent compared to global aquaculture production in 2019 (<u>FAO/FishStat1, 2021</u>).

Species group		Finfish Crustacean										
Continent	Africa	Americas	Asia	Europe	Oceania	World	Africa	Americas	Asia	Europe	Oceania	World
				Inland aq	uaculture	productio	n					
Species group	1,966	1,151	44,713	536	5	48,370	0	73	4,108	0	0	4,181
Total inland aquaculture	1,966	1,224	49,572	536	5	53,303	1,966	1,224	49,572	536	5	53,303
Species group % of total	100%	94%	90%	100%	98%	91%	0%	6%	8%	0%	2%	8%
				Marine aq	uaculture	productio	n					
Species group	299	1,152	4,369	2,052	84	7,957	5	1,075	5,213	0	6	6,301
Total marine aquaculture	311	2,956	25,889	2,700	204	32,060	311	2,956	25,889	2,700	204	32,060
Species group % of total	96%	39%	17%	76%	41%	25%	2%	36%	20%	0%	3%	20%
				Global aq	uaculture	prodction						
Species group	2,265	2,303	49,082	2,588	89	56,327	5	1,148	9,321	1	7	10,481
Total aquaculture	2,277	4,179	75,461	3,236	209	85,363	2,277	4,179	75,461	3,236	209	85,363
Species group % of total	99%	55%	65%	80%	43%	66%	0%	27%	12%	0%	3%	12%
Inland and marine aquaculture as % of production by continent												
% inland aquaculture	87%	50%	91%	21%	5%	86%	0%	6%	44%	36%	1%	40%
% marine aquaculture	13%	50%	9%	79%	95%	14%	100%	94%	56%	64%	99%	60%

192

193 Many factors are affecting crustacean production, including climate, technology, disease, natural disaster, 194 pandemic, economy, etc., but these events are occurring irregularly, and have a minor effect on global 195 production. Historically, disease outbreaks have been causing severe losses in crustacean production and 196 can be traced in production statistics of major shrimp producing countries, showing large drops in regional 197 production. For example, white spot syndrome virus (WSSV) and early mortality syndrome (EMS = AHPND) 198 outbreaks in major shrimp producing countries caused significant production declines and hence affected 199 the year-on-year growth rate globally and in Asia (Shinn et al., 2018). On the other hand, global crustacean 200 production increases, as shown by consistent positive AAGR values (Table 2), Nevertheless, AAGRs would have been higher, provided disease related losses would have been lower (Stentiford et al., 2012). 201



Figure 2

The percentage year-on-year change in the growth of Asian and global shrimp production (Shinn et al., 2018).

¹ Data from FAO FishStatJ (2017) and national feed sale figures (where available) are used.

² AHPND = acute hepatopancreatic necrosis disease, EHP = Enterocytozoon hepatopenaei, IHHNV = infectious hypodermal and haematopoietic necrosis virus, TSV = Taura syndrome virus, WSSV = white-spot syndrome virus, YHV = yellow head virus.

204 2.1.1.2 Main culture species

In 2019, there were 13 finfish species with a production above 1 Mt (Table 4), representing 71% of the global finfish production. Of these 13 species seven were carps, with a combined production of 26 Mt in 2019. The share of these carp species to global finfish production declined from 60% in 2000 to 46% in 2019. This decline was to a large extend compensated by increased production of tilapias and catfishes, whose contribution to global production increased with 5% and 6%, respectively, between 2000 and 2019. The percentage contributions of Atlantic salmon and milkfish to global finfish production increased 1% between 2000 and 2019, providing 5 and 3%, respectively, of global finfish production in 2019.

212 Table 4: Annual production of finfish species with a production above 1 million MT yr⁻¹
 213 (<u>FAO/FishStatJ, 2021</u>).

		20	000	2019			
Species (> 1 mil	lion MT yr ⁻¹)	production (Mt yr ⁻¹)	% global production	production (Mt yr ⁻¹)	% global production		
		Carps		•			
Grass carp	Ctenopharyngodon idellus	3.0		5.7			
Silver carp	Hypoththalmichthys molitrix	3.0		4.8	シ		
Common carp	Cyprinus carpio	2.4		4.4			
Catla	Catla catla	0.6		3.3			
Bighead carp	Hypophthalmichthys nobilis	1.4		3.1			
Crucian carps	Carassius spp.	1.2		2.8			
Rohu	Labeo rohita	0.7		2.0			
Total		12.4	60%	26.1	46%		
		Tilapias					
Nile tilapia	Oreochromis niloticus	1.0		4.6			
Tilapia nei	Oreochromis spp.	0.1		1.1			
Total		1.1	5%	5.7	10%		
		Catfishes					
Striped catfish	Pangasionodon hypophthalmus	0.1		2.6			
Torpedo-shaped catfishes nei	Clarias spp.	0.0		1.3			
Total		0.2	1%	3.9	7%		
		Salmonids					
Atlantic salmon	Salmo salar	0.9	4%	2.6	5%		
		Milkfish			-		
Milkfish	Chanos chanos	0.5	2%	1.5	3%		
Total		15.0	72%	39.9	71%		
Global finfish n	roduction	20.8		56.3			

214

In 1950, when aquaculture was still in its infancy, freshwater species provided 78% of total production,
diadromous species 21% and marine species 1%. By 2000, the share of freshwater species to total
production increased to 84%, diadromous species dropped to 11% and marine species increased to 5%.
After 2000, the contributions of marine fishes increased 1%, mainly compensating a decline in diadromous
fishes (Table 5; (FAO/FishStatJ, 2021).

220 *Table 5:*

Percentage contribution of freshwater, diadromous and marine finfish species to global aquaculture production (<u>FAO/FishStatJ, 2021</u>).

	1950	2000	2019
Freshwater fishes	78	84	84
Diadromous fishes	21	11	10
Marine fishes	1	5	6

222 223

221

225 In 2019 there were 4 crustacean species with a production above 0.75 Mt, together providing 87% of the 226 global crustacean aquaculture production (Table 6). This share most likely will continue to increase slowly 227 during the next decade. Whiteleg shrimp (Penaeus vannamei) alone, with a production of 5.4 Mt provides 228 more than half of the global crustacean production through aquaculture. Red swamp crayfish (Procambarus 229 clarkii), the majority of which is produced in rice-crayfish farming systems in China (China National 230 Statistics 2020 provided by Jie Huang, NACA), comes in second with a production of 2.2 Mt in 2019 231 (FAO/FishStatJ, 2021). Within less than 20 year, culture of this species exploded, from 0.5% of the global 232 crustacean production in 2000 to 21% in 2019, due to market promotion followed by consumer preference (Wang et al., 2018). Chinese mitten crab (Eriocheir sinensis) comes third with a production of 0.78 Mt, 233 234 followed by giant tiger prawn (Penaeus monodon) with 0.77 Mt, each contributing 7% to the global 235 crustacean production in 2019. Mitten crab culture is practiced mainly in China. For juveniles the principal 236 culture system is rice-crab polyculture, while grow-out is mainly done in ponds, all or not in polyculture 237 with other species (Cheng et al., 2018). Between 2000 and 2019, whiteleg shrimp and giant tiger prawn 238 swapped leading positions as most important crustacean aquaculture species. However, large 239 improvements were made through selection in growth and disease resistance of giant tiger prawn. Its high 240 price and the possibility to stock the new strains at high density in ponds triggers speculation about a possible comeback during the next decade (The Shrimp Blog | Shrimp Insights, may 2021). 241

242 243

Table 6:Crustacean species with a production above 0.75 million metric tonne (Mt) in 2019
(FAO/FishStatJ, 2021), and their contribution to global crustacean production.

		200	00	2019			
English name	Coiontifio no mo	production	% global	production	% global		
English name	Scientific name	('000 MT yr⁻¹)	production	('000 MT yr ⁻¹)	production		
Whiteleg shrimp	Penaeus vannamei	155	9	5,446	52		
Red swamp crayfish	Procambarus clarkii	8	0.5	2,162	20.6		
Chinese mitten crab	Eriocheir sinensis	203	12	779	7		
Giant tiger prawn	Penaeus monodon	631	37	774	7		
Total		996	59	9,162	87		
	Global production	1,691		10,481			

244 245

246 2.1.2 Extractive species

247 2.1.2.1 Production overview

248 The share of molluscs to the global animal production through aquaculture dropped from 30% in 2000 to 21% in 2019. The AAGR of molluscs is with 3.5 and 2.7% for the periods 2000-2010 and 2010-2019, 249 250 respectively, much lower than the global aquaculture AAGRs of 6.1 and 4.9% over the same periods (Table 251 2). Of the global mollusc production in 2019, only 1% was produced in inland waters. In marine areas, molluscs provide 54% of the aquaculture production in marine areas. In Africa molluscs production is 252 253 negligible small. In Oceania, molluscs production is also small, but responsible for 56% of the total 254 aquaculture production on the continent. In the Americas, Europe and Asia, molluscs aquaculture is 255 important, providing 17, 20 and 21% of the total aquaculture production on the respective continents 256 (Table 7).

Similar to molluscs, 99% of aquatic algae are produced in marine areas. The vast majority of aquatic plants are produced in Asia, with only 0.5% of the global production produced outside Asia. Nevertheless, all other continents are experimenting with algae culture and report productions (Table 7). Interest in seaweed (= macroalgae) for improved nutrition, industrial use, and ecosystem services, grew globally during the last decades, beyond the main producing countries China, Japan, Korea, and parts of South America (<u>Buschmann et al., 2017</u>). Overall, seaweed aquaculture has not been analyzed in depth (<u>Chopin and</u> <u>Tacon, 2020</u>).

264 FAO statistics related to the production of aquatic plants and algae shows production tripled from 10 Mt of 265 wet biomass in 2000 to 35 Mt in 2019. Table 9 summarizes production, culture system and use of the main 266 species of seaweed. Seaweeds are special, considering ca. 31 - 38% of the global seaweed production is 267 consumed directly as food (Naylor et al., 2021). Today, the brown seaweed Saccharina japonica (wakame) 268 consumed for food and alginate production, and the red algae *Euchema* spp. for the carrageenan industry, are the 1st and 2nd most productive aquaculture species worldwide (Buschmann et al., 2017). The majority 269 270 of seaweed biomass is used by the industry sector as polysaccharide additives and functional 271 food ingredients, and by the non-food sector as hydrocolloid products in nutraceuticals, 272 pharmaceuticals and cosmetics, and to a lesser extent as fertilizers, feed ingredients, biofuels, bioplastics, and other industrial outputs (Naylor et al., 2021). 273

- 274
- 275Table 7:Molluscs and aquatic plants production (thousand metric ton) through aquaculture by continent276in 2019. Mollusc production is compared to global animal production through aquaculture.277Aquatic algae production is compared to the global aquaculture production, including algae278(FAO/FishStatJ, 2021). When cells are empty no values have been reported.

9

279

Species group		Molluscs					Aquatic algae					
Continent	Africa	Americas	Asia	Europe	Oceania	World	Africa	Americas	Asia	Europe	Oceania	World
				Inland a	quaculture	e productio	n					
Species group			201			201	0.4	1	55	0.4		56
Total inland aquaculture	1,966	1,224	49,572	536	5	53,303	1,966	1,225	49,627	536	5	53,359
Species group % of total			0.4%			0.4%	0.0%	0.1%	0.1%	0.1%		0.1%
	Marine aquaculture production											
Species group	7	728	15,885	643	113	17,376	118	23	34,513	11	14	34,679
Total marine aquaculture	311	2,956	25,889	2,700	204	32,060	429	2,979	60,402	2,711	218	66,739
Species group % of total	2%	25%	61%	24%	56%	54%	27%	1%	57%	0%	6%	52%
				Global a	quaculture	e productio	n					
Species group	7	728	16,086	643	113	17,577	118	24	34,568	11	14	34,736
Total aquaculture	2,277	4,179	75,461	3,236	209	85,363	2,395	4,203	110,029	3,248	223	120,098
Species group % of total	0%	17%	21%	20%	54%	21%	5%	1%	31%	0%	6%	29%
	Inland and marine aquaculture as % of production by continent											
% inland aquaculture			1%			1%	0.3%	4%	0.2%	3%		0.2%
% marine aquaculture	100%	100%	99%	100%	100%	99%	99.7%	96%	99.8%	97%	100%	99.8%

280 281

282 2.1.2.2 Main culture species

Cupped oysters and Japanese carpet shell dominate molluscs production, followed by scallops and sea mussels. Together these 4 culture groups provide 70% of the global mollusc production (Table 8). The highest production is for cupped oysters, responsible for 30% of the global mollusc production. The highest production increase was for Japanese carpet shell, raising production 170% between 2000 and 2019, to provide 23% of the global molluscs production in 2019.

288 Table 8: 289 Mollusc species with an production above 1 Mt in 2019 (<u>FAO/FishStatJ, 2021</u>), and their contribution to global crustacean production.

	20	00	2019		
English name	Scientific name	production	% global	production	% global
English harne	Scientific fiame	('000 MT yr ⁻¹)	production	('000 MT yr ⁻¹)	production
Cupped oysters nei	Crassostrea spp.	2,923	27	5,265	30
Japanese carpet shell	Ruditapes philippinarus	1,504	14	4,028	23
Scallops nei	Pectinidae	811	7	1,828	10
Sea mussels nei	Mitylidae	720	7	1,116	6
T	otal	5,958	55	12,237	70
Global p	roduction	10,866		17,577	

Three brown algae and 4 red algae species have an annual production above 1 Mt (Table 9). Together they represented 96% of the aquatic plants production in 2019. The most important culture species are the brown algae Japanese kelp (*Laminaria japonica*) providing 39% of global production, and the red algae *Euchema* spp. and *Gracilaria* spp. providing 28 and 10%, respectively, of global production.

296 It has been postulated that seaweeds could substitute some terrestrial crops and animal production in 297 protein, fat (omega 3) and energy intake, alleviating pressure on freshwater and land use and impact on 298 biodiversity, but there is little evidence to date that seaweeds will contribute substantially to human 299 macronutrient intake in the future (Wells et al., 2017). On the other hand, seaweed farming is widely 300 recognized for its ecosystem services beyond the provision of food and feed, yet producers have not been 301 able to capture this value in financial returns (Chopin and Tacon, 2020). Bioremediation is one of the main 302 services propagated and studies at large scale seaweed farming areas indicate these organisms are 303 effective in reducing nitrogen levels, controlling phytoplankton blooms, and limiting the frequency of toxic 304 algal blooms (Xiao et al., 2017; Yang et al., 2015). In addition, large-scale aquaculture of seaweed 305 positively regulates and improves environmental conditions in coastal ecosystems (Xiao et al., 2021). 306 However, the effectiveness of impact of ecosystem services provided by seaweed farming, still requires 307 attention across cultured systems, seasons, and scales.

308Table 9:The production, culture system and use of the major seaweeds cultured in 2019, their309production system, and use for food and non-food purposes. Species listed represent 99% of310the global seaweed production. Species with a production above 1 Mt are shaded grey311(FAO/FishStatJ, 2021).

	Culture System		Production	Use	
Cultured Seaweed	Primary	Secondary	Ton (fresh)	% used as food	Other uses
Brown Algae					
Alaria esculenta	Suspended long-lines		105	100	
Laminaria japonica	Suspended long-lines		12,273,519	50	Alginate industry
Macrocystis pyrifera	Suspended long-lines		2	0	
Nemacystus decipiens	?		20	100	
Saccharina latissima	Suspended long-lines		229	20	Alginate industry
Sargassum fusiforme	Suspended long-lines		303,797	100	
Undaria pinnatifida	Suspended long-lines		2,563,477	100	
Other Phaeophyceae	?		1,252,264	unknown	
TOTAL			16,393,413	55	
Red Algae					
Eucheuma denticulatum	Suspended long-lines	Lines attached to the bottom	179,360	0	
Eucheuma spp.	Suspended long-lines	Lines attached to the bottom in shallow waters	9,817,689	0	
Gracilaria spp.	Suspended long-lines		3,638,554	1	Agar industry
Gracilaria gracilis	Wild harvesting		273	0	Agar industry
Gracilaria verrucosa	Bottom culture		1006	0	Agar Industry
Kappaphyccus alvarezii	Suspended long-lines		1,625,164	0	Carrageenan industry
Porphyra spp.	Suspended nets	Intertidal based nets	2,123,040	100	
Porphyra tenera			861,083	100	
TOTAL			18,246,169	17	
Green Algae					
Capsosiphon fulvescens	?		3,386	100	
Caulerpa spp.	ponds		1090	100	
Chlorella vulgaris	Bioreactors		5	0	Health Food
Codium fragile	ponds		3,258	100	
Dunaliella salina	Bioreactors		0	0	Carotene production
Enteromorpha prolifera	Suspended Long-lines		0	100	
Haematococcus pluvialis	Biorectors		242	0	Carotene Production
Monostroma nitidum	Suspended long-lines		6,321	100	
TOTAL			14,302	98	
GRAND TOTAL			34,653,884	35	
FAO Reported prod	uction in 2019 (F	ishStat1)	34,735,590		

312 313

314 2.2 Aquaculture systems

315 2.2.1 Fed systems for finfish

316 Forty years ago it was still common practice to use locally available crop wastes, manure, waste water, or 317 grains as nutrient sources to stimulate fish production in aquaculture ponds (Hickling, 1962; Huet, 1986). 318 Today, the role of locally available ingredients in many integrated agriculture-aquaculture farming systems 319 has been replaced by pelleted feed (Tacon, 2020). Because pelleted feeds are produced off-farm, farmers 320 were no longer dependent on a limited supply of on-farm or locally nutrients, allowing them to intensify. 321 With this feed-driven intensification, the contribution of these mixed farming systems to total finfish 322 production declined (Edwards, 2015). Nevertheless, still numerous small-scale farmers depend upon 323 integrated pond farming systems, not only for fish, but also for water, and to improve food security (Ahmed 324 et al., 2014).

Until recently, in developing countries, a distinction was made between small-scale and industrial fish farming. Small-scale aquaculture provided a source of animal protein for home consumption and/or surplus income from fish sold locally to poor and food insecure households (<u>Ahmed and Lorica, 2002</u>). This notion required governments and donors to propagate small-scale aquaculture to reduce hunger and improve food security, and to support education in aquaculture. The knowledge level of farmers improved, facilitating the development of a largely overlooked 'missing middle' of commercially oriented fish farmers.

These middle farmers were reacting to increased demand for fish and buying power within their Mainly low value species, including carps, tilapias, and catfishes are raised by these 'middle' farmers (<u>Belton et al., 2018</u>). As such, aquaculture contributed to rural development and poverty alleviation, making aquaculture an integral part of the rural economy.

The mix of small-, middle-, and large-scale commercial farmers has made finfish production in ponds highly diverse. Until early 2000, production ranged between 50 and 100 000 kg/ha (Table 10). In stocked ponds receiving no fertilizers and feed, production levels of 50-500 kg/ha were reached. With a combination of fertilizers and supplemental feeds, production increased to 1 000 - 4 000 kg/ha, which could be further increased to 10 000 kg/ha using well-formulated feeds and aeration. Without water exchange, production up to 10 000 kg/ha was possible, with dissolved oxygen availability being the principal factor limiting production.

344

Table 10: Aquaculture production at increasing levels of input for pond-raised channel catfish and tilapia in 1985 until early 2000. (Modified from <u>Boyd (1990)</u> and <u>Verdegem et al. (2006)</u> in(<u>Tucker and Hargreaves, 2012</u>).

	Annual prod	uction (kg/ha)	
Management input	Channel catfish	Tilapia	Limiting factor
Stocking only	50-100	200-500	primary productivity
Stocking, fertilization	200-300	1,000-3,000	primary productivity
Stocking, fertilization, supplemental feeding	500-1,000	3,000-4,000	dissolved oxygen
Stocking, feeding	1,000–2,000	3,000-4,000	dissolved oxygen
Stocking, feeding, emergency aeration	4,000-6,000	4,000-6,000	dissolved oxygen
Stocking, feeding, continuous aeration	6,000–10,000	6,000–10,000	dissolved oxygen, metabolites
Stocking, feeding, continuous aeration, water exchange	10,000-20,000	15,000-35,000	metabolites
Stocking, feeding, continuous aeration, intensive mixing		20,000-100,000	metabolites, suspended solids

345

346 Production levels 20 000 – 35 000 up to 100 000 kg/ha become possible, when removing metabolites 347 (e.g. NH₄, CO₂) and suspended solids (Table 4). The latter can be done through water exchange or by 348 stimulating in-pond water purification processes, as done in partitioned ponds, in-pond raceways, or split 349 ponds. In these systems, fish are raised in high density in 5 – 25% of the pond area. The remaining pond 350 area mainly acts as an algal reactor, where algae remove metabolites and provide oxygen. The higher the 351 primary productivity, the higher the capacity of the pond to hold fish. If the primary production is raised 352 by a factor 2 to 4, the production capacity of the pond raises with the same factor. These systems work 353 well, provided the algal density is controlled to avoid dissolved oxygen depletion due to algal die-off 354 (Cremer and Chappell, 2014; Tucker and Hargreaves, 2012). A technique used in the past, but with little 355 follow up today, is confinement of fed fish in in-pond cages, with free roaming fish feeding on natural food 356 between cages fed pelleted feed. The increase in primary production was small, raising total production by 357 ca. 25%. Another in-pond water purification technique is the creation of biofloc through intensive aeration 358 and water agitation. Bioflocs are a mix of autotrophic and heterotrophic bacteria that purify the water and 359 can be eaten by filter feeders (e.g. tilapia) improving the feed utilization efficiency (Avnimelech, 2007; 360 Dauda, 2020).

Since the first use of fish feeds, the quality of pelleted feed continually improved (Edwards, 2015), due to better processing (extrusion, floating or sinking pellets, higher water stability), formulation (novel ingredients, better balance of macro- and micro-nutrients) and use of additives (prebiotics, probiotics, enzymes). Better feed quality improves the feed utilization efficiency and reduces waste accumulation, leading to 15 – 40% higher production levels today than indicated in Table 4 under similar management conditions (Boyd et al., 2020).

³⁴² 343

367 With the development of feeds that directly address the nutrient requirements of culture species and 368 availability of efficient electric powered aerators (Boyd et al., 2020), the self-purifying capacity of ponds 369 improved (Boyd and Chainark, 2009) allowing increase of feed inputs. This concurred with improvements 370 in nutrient utilization efficiency, and less waste accumulation and discharge per kg fish produced. 371 Nevertheless, due to the high culture intensity, more nutrients are produced than the pond can handle. An important constraint is that fish waste resulting from pelleted feed is nutrient rich (e.g. nitrogen, 372 373 phosphorous) and energy poor (e.g. carbon), causing microorganisms in the pond lacking energy to 374 mineralize the waste. Raising the C:N ratio of the nutrient input to aquatic systems above 12 by providing 375 more carbohydrates, fish performance (e.g. growth, survival, FCR) and water quality improves, in systems 376 ranging from extensive to intensive, the latter including biofloc systems (Asaduzzaman et al., 2009; Kabir 377 et al., 2019). A constraint is that by raising the carbon input, the carbon emission from the pond increases 378 (Tinh et al., 2021). Therefore, Kabir et al. (2020) replaced in pelleted feed easily digestible starch with 379 fibers which are difficult to digest by the fish. A large fraction of these fibres will enter the pond through 380 the faeces, where microorganisms can break them down to get energy. As a result, more nutrients are 381 assimilated in the food web, providing natural food. Using this concept, up to 74% of the nitrogen supplied 382 with the feed was harvested in fish biomass, without a need to add extra carbon to the system. In 383 consequence, CO₂ emissions also declined. This nutritious pond concept (<u>loffre and Verdegem, 2019</u>) can 384 be applied with herbivorous and omnivorous species in fed and/or aerated ponds with a biomass up to 15 385 000 kg/ha at harvest.

386 To intensify further, wastes should be either discharged, recycled, or purified. Discharging nutrients without 387 treatment causes environmental damage (e.g. eutrophication, biodiversity loss) (Boyd et al., 2007), and 388 this practice should be abandoned. Options are to collect wastes and turn them into fertilizers or biogas. 389 Wastes can also be (partially) recuperated through integrated multi-trophic aquaculture, or treated in 390 recirculation systems (sections below). As an industry, the combination of all these technologies made 391 aquaculture more environmentally sustainable and resource efficient. In addition, these mature 392 technologies can be adjusted to other geographies, facilitating the development of sustainable aquaculture 393 outside Asia.

It should be noted that ponds are integrated in the landscape, playing a key role in trapping nutrients from run-off, and the apparent nutrient use efficiency can be higher than for other animal production systems. A broader analysis of the reactive nitrogen (N*r*) flow in Chinese inland aquaculture ecosystems suggests 72% of the feed nitrogen is retained in aquatic products (Luo et al., 2018). This is a very high efficiency, and, in doing so, more focus on aquaculture can make food production systems more resilient to climate change.

400 2.2.2 Crustacean aquaculture systems

Limiting factors that influence finfish culture (Table 4) also affect crustacean production. One important difference is that most crustaceans live on the bottom. Therefore it is important to maintain the bottom clean and with sufficient oxygen. When wastes accumulating at the bottom are not regularly removed, this restricts the culture capacity for crustaceans, resulting on average in a lower production output than with finfish for similarly designed and managed finfish ponds, and thus higher operational costs. In addition, crustaceans are vulnerable to predation and reduced feed intake when molting.

407 Shrimp aquaculture started with stocking post larvae (PL) that were caught in the coastal zone in ponds 408 with tidal water exchange. The latter helped to maintain water quality and provided nutrients to the system. 409 The farming system gradually improved. Hatchery-reared PL replaced wild caught PL and commercial feed 410 was applied during the last month of culture. Systems were extensive, stocking 2 - 15 PL/m². Semi-411 intensive culture systems use emergency aeration, pelleted feed, and exchange water daily, especially during the last months of culture, stocking 25-30 PL/m² and producing 3 – 6 tonne/ha. Intensive cultures 412 413 start with stocking densities of $30-50 \text{ PL/m}^2$ aiming to produce 10 tonne/ha. This goes up to 300 - 800414 PL/m^2 in super-intensive systems to produce 35 - 110 tonne/ha.

415 The more intensive, the more aeration, water exchange, probiotics, and feed additives are used to maintain

a healthy culture environment. Production is further enhanced by intermediate partial harvests, allowing

- to produce more while staying within the carrying capacity of the farming system, providing more frequent
- 418 income.

419 Losses due to disease in shrimp farming are high (Shinn et al., 2018; Stentiford et al., 2012). Various

- 420 approaches to minimize the impact of disease on production are possible. In Latin America, the aim is to
- 421 develop farming systems resilient to disease and to cope with pathogens present in the production 422 environment (<u>Alday-Sanz, 2018</u>). Large semi-intensively operated ponds are used for grow-out, focusing
- 423 on maintaining a healthy culture environment.
- Another approach to keep the pathogen pressure low is polyculture of shrimp and finfish, either in crop rotation or in co-culture (<u>Paclibare et al., 1998</u>). This practice makes shrimp farming more sustainable (<u>Yi</u> and <u>Fitzsimmons, 2004</u>) by reducing the environmental impact, and reducing the incidence of shrimp disease (<u>Halim and Juanri, 2016</u>; <u>Martínez-Porchas et al., 2010</u>). One method is to stock fish (e.g. tilapia) in a reservoir pond and to circulate water between the reservoir and adjacent shrimp ponds. Antimicrobial peptides in the fish skin kill shrimp pathogens, keeping pathogen pressure of bacteria (<u>Masso-Silva and</u> <u>Diamond, 2014</u>) and viruses low. The latter approach was applied in a tilapia-shrimp intensive polyculture
- 430 <u>Diamona, 2014</u>) and viruses low. The latter app
 - 431 systems in Indonesia (Figure 3).
 - 432



Example of an integrated intensive tilapia-shrimp polyculture system.

The approach is flexible, as the water exchange between the fish pond and polyculture pond can vary, as well as the stocking density of tilapia in the polyculture pond.

In case disease occurs, it is important to make sure pathogens are eliminated from waste leaving the farm.

unpublished data, PT. AWS, Indonesia, 2011

434

435 The use of specific pathogen free (SPF) or specific pathogen resistant (SPR) shrimp stocks allowed 436 hatcheries and broodstock multiplication centers to upgrade biosecurity, benefiting the downstream grow-437 out farming sector. In addition, large corporate producers incorporated this aspect into the quality 438 assurance system linked to their biosecurity and operation's management. Every input into the hatchery 439 or farm requires quality control approval as part of the quality assurance system. This results in higher 440 profits and sustainability, and could be applied more widely, to raise the efficiency across the shrimp 441 industry. The 'vannamei PL efficiency index' calculates the amount (tonne) of shrimp produced per million 442 PL stocked by country. The index gives a broad indication of disease related losses. Whereas strong-443 producing countries have an index of 9 - 11 tonne/million PL, the index can be close to 1 in poor-performing 444 countries, indicating that large improvements across the shrimp industry are still possible (Merican, 2021).

Technological solutions are also proposed. One approach is to apply a combination of using (i) small, easy to operate, grow-out ponds, (ii) high on-farm aeration and energy capacity, (iii) a shrimp 'toilet' at the center of each pond, and (iv) a large reservoir area for water exchange, taking 60% of the farm area and leaving maximum 40% for grow-out (Kawabigashi 2018) The goal is to maintain a clean bottom environment, while minimizing off-farm water exchange during culture for biosecurity and operating small, easy to manage, super-intensive grow-out ponds. Different mixes of physical and chemical treatment methods are today commonly applied in shrimp hatcheries and grow-out operations using partial or complete combinations of sedimentation, filtration, foam fractionation, ozonation, and UV irradiation.

454 Another important technological improvement is the development of precision hardware and software 455 allowing hatcheries and farms to collect reliable data (e.g. PL or larval counting with >95% accuracy) and 456 get feedback through artificial intelligence and Internet of Things. A similar development is ongoing in 457 feeding technology. Commercial feeds represent 60% of the operating cost of shrimp farming. The use of 458 automatic feeders, reacting to noise made by foraging shrimp, are effective in reducing feeding costs and 459 improving efficient and profitable nutrient use. This technology is presently spreading quickly through the 460 industry. Joint efforts between feed manufacturers and companies making automated feeders to adjust 461 the physical properties of manufactured aquafeeds to automatic feeder design and operation are ongoing, 462 and will lead to further improvements in feeding efficiency (Molina and Espinoza, 2018).

463 2.2.3 Extractive systems

464 2.2.3.1 Molluscs

While there are common and general methods and gear for shellfish grow-out, there is some regional 465 466 specificity with regard to methodologies. Many innovations are shared between growers and incorporated 467 into local practices. Bottom culture employs direct planting, mesh bottom culture bags, rack and bag 468 systems (oysters and clams), and off-bottom culture systems using primarily long-line and raft systems 469 with hanging lines (mussels, scallops). Offshore aquaculture requires specialized gear that is technically and economically viable, can withstand the challenges of the offshore environment, and meet government 470 471 regulations. There are successful offshore aquaculture systems in operation currently, but they are few. 472 Many are still experimental or struggling to obtain the necessary permits to allow commercial expansion.

473 Reliance on wild seed/spat for culture of bivalve molluscs is still very high for many regions and species 474 globally. Hatchery design and technology have made major advances during the last few decades in 475 conditioning, spawning, larval care, and setting, with higher survival of the animals seen as this occurred. 476 As noted by Sarkis et al. (2021), they vary with site characteristics, target species, production level, culture 477 methodologies, and available funds, and range from small family operations to large corporate multi-478 species facilities with automated systems. Phytoplankton production in hatcheries has similarly advanced 479 with computer-aided monitoring and metering of feed to larval shellfish, again enhancing survival and 480 growth. Development of improved setting procedures and equipment have allowed growers to produce 481 seed aimed at their specific needs, and advances in large-scale setting and planting, especially of oysters, 482 was a direct result of more effective handling of materials.

483 Most molluscan shellfish in aquaculture such as oysters, clams, scallops and mussels are filter-feeders 484 that do not require artificial input of feed. Farming of these species has little impact on the environment 485 and is expected to expand significantly in the future (Shumway, 2011). Molluscan shellfish are cultured in 486 open waters and are subjected to wild fluctuations in environmental parameters. Molluscs have high 487 fecundity and a Type III survivorship and, even in the adult stage, they are prone to mass mortality 488 triggered by environmental stress and diseases. Climate change, e.g. warming and acidification of the 489 ocean, pose a great challenge to molluscan aquaculture. Most molluscan shellfish in aquaculture have little 490 or no history of domestication. Selective breeding and advanced genetic improvements are therefore 491 needed to enhance production efficiency and resilience of molluscan shellfish.

Selective breeding has contributed to shellfish aquaculture through the development of disease-resistant and fast-growing strains, and varieties with unique shell colors (<u>Guo, 2021</u>). Sterile triploids that grow faster and maintain meat quality during their spawning season have become an important of oyster farming. Hybridization has also proven to be useful in genetic improvement of molluscs. In China and elsewhere, over 30 molluscan species have been subjected to some genetic improvement, although the level of breeding is far inadequate for what the shellfish aquaculture industry needs.

498 Long-term and well-maintained shellfish breeding programs are few, and genetic improvements are mostly 499 moderate. While traditional selective breeding has yet to reach adequate levels in most molluscs, new 500 breeding technologies are posed to make a great impact on aquaculture (Stokstad, 2020). Genomic 501 selection that has demonstrated its power in agriculture crops and livestock, promises to accelerate genetic 502 improvement of molluscan shellfish. Genomic selection is more effective and can eliminate the need for 503 expensive phenotyping. Genomes have been sequenced for many cultured molluscs, and high-throughput 504 genotyping platforms such as single-nucleotide polymorphism (SNP) chip are being developed in several 505 species for genomic selection. Gene-editing through CRISPR/cas9 offers the opportunity of modifying 506 target genes in a way that mimics natural mutation, an approach that is fundamentally different from the introduction of foreign genes in genetic engineering. While these new technologies are exciting, they have 507 508 to be rigorously evaluated for shellfish aquaculture following proper regulations and with the support of 509 the shellfish industry.

510

511 2.2.3.2 Seaweed

512 During the past 300 years, and particularly since the second half of the 20 century, mariculture of seaweed 513 became common, starting with culturing wild seaweed (Buschmann et al., 2017). The discovery that the 514 commonly grown Pyropia life-history included heteromorphic life stages identified previously as two 515 different species allowed the development of a whole farming industry that now provides nori globally. 516 Around the same time, the introduction of the kelp Saccharina japonica (formerly Laminaria japonica, 517 kombu) in China, allowed the change from simply harvesting the wild seaweed stands to culture of 518 selectedg strains, reproduced under controlled conditions. From this point, other countries such as Korea also started culturing seaweed. Soon, seaweed mariculture bypassed the amounts of seaweed collected in 519 520 the wild. In contrast to other aquaculture commodities, the amount of seaweed collected in the wild is today negligible compared to the amount of seaweed cultured (Chopin and Tacon, 2020; Naylor et al., 521 522 2021). Nevertheless, considering the large number of seaweed species, and with 7 species to provide 97% 523 of the global production of aquatic plants, there remain many species today to be tested for culture. In 524 consequence, the full potential of seaweed mariculture remains largely unknown, and requires further 525 research (Hafting et al., 2015).

Algal productivity depends primarily on light and nutrient availability, with nitrogen being the limiting 526 527 nutrient in the marine environment. Important modulating factors include temperature, salinity, pH, and 528 other environmental factors affecting seaweed metabolism (Santelices, 1999). As light decreases rapidly 529 with depth, the development of floating near-surface cultivation technologies gave a major boost to the 530 expansion of seaweed culture, allowing to culture in regions further offshore. The majority of seaweeds 531 today use near-surface cultivation techniques (Table 9). The growth in production of Pyropia and 532 Eucheuma/Kappaphyccus illustrates this point (López-Vivas et al., 2015). In addition , the industry seeks 533 to raise value by developing highly productive strains and through novel processing technologies both for 534 food and phycocolloid extraction (Hwang and Park, 2020). Nevertheless, domestication of seaweeds demands better insight into the ecological and genetic diversity of wild populations and the impact of 535 536 cultivars on the production environment (Valero et al., 2017). Experience with seaweed farming, including 537 how to make optimal use of the prevailing environmental conditions at the production sites, is largely 538 missing in occidental countries, making capacity building a prerequisite to successful development of 539 seaweed farming (Santelices, 1999).

540

541 2.2.4 Inland and marine cage aquaculture

542 With the exception of extensive cage culture of filter feeding species (e.g. silver and bighead carp, milkfish) 543 in eutrophic lakes (<u>Delmendo and Gedney, 1976; Husen et al., 2012</u>), cage aquaculture is intensive, relying 544 on pelleted feed. Fish can be maintained in high density, as feed wastes are flushed out from the pond 545 with exchange water. If the waste loading is higher than the lake can absorb fish kills might occur due to 546 oxygen depletion. Therefore, limits must be set to the total biomass produced in public waterbodies.

547 Models exist to indicate safe limits to production (<u>David et al., 2015</u>). Pollution in marine areas from cage 548 aquaculture is also a concern, especially in shallow, partially enclosed areas.

In 2019, 16% of the global finfish production was produced in marine or coastal environments, of which 549 550 nearly 1/3rd was Atlantic salmon (Table 2 and 3). High value and high profit margins give Atlantic salmon, 551 which is the third most valuable aquaculture finfish species produced today, room to innovate, including 552 making larger and stronger cages which can be deployed offshore, away from heavily used coastal zones 553 (Chu et al., 2020). A downside is the high capital and production costs involved (Jansen et al., 2016), 554 which also holds for other species cultured at sea. This partially explains the slow growth of marine cage 555 aquaculture, in spite of the fact that from an environmental point of view there is ample of room for 556 development of marine cage aquaculture (Gentry et al., 2017). Another reason is that herbivorous or 557 omnivorous species produced inland are cheaper than marine species which are mostly carnivores and 558 more expensive to produce. For now, cheaper herbivorous and omnivorous fish species are more widely 559 accessible and contribute significantly to food security and reduced poverty reduction. This trend will not change within the near future (Belton et al., 2018; Belton et al., 2020), but might change over the long 560 term. Considering the projected increase in global income, Costello et al. (2020) predicts a 36-74% 561 increase in production of marine aquaculture by 2050, when the majority of people will be middle class. 562

563 Coastal areas for shrimp pond development are limited while the sector aims to expand further. One option 564 is cage farming. Crustaceans especially shrimps are difficult to culture in marine cages, mainly due to 565 cannibalism during molting. Shrimp culture in cages, either as shrimp – seaweed polyculture (<u>Lombardi et</u> 566 al., 2006) or monoculture at varying stocking densities (<u>Cuvin-Aralar et al., 2009</u>) looked economically 567 feasible, also at high stocking density. Biosecurity is, however, a problem in marine shrimp culture at sea, 568 as is pollution control. The latter might be partially controlled by the implementation of integrated 569 multitrophic aquaculture (section 2.4.6).

570 2.2.5 Recirculation aquaculture systems (RAS)

571 The term Recirculating Aquaculture Systems (RAS) is not well defined. The key element is that RAS use 572 significantly less water than traditional systems by fully or partially purifying and reusing culture water. 573 They range from semi-intensive open systems to fully recirculating indoor systems and are used for 574 producing a broad range of freshwater and saline species. While mainly finfish are produced in RAS, also 575 crustaceans, molluscs, and aquatic plants can be produced, the latter mainly for hatchery and nursery 576 purposes. The simplest RAS remove suspended solids (faeces and uneaten feed) from the recirculation 577 loop by sedimentation or mechanical filtration and aeration (e.g. airlifts), while higher intensity RAS apply biofiltration and more advanced treatment technologies such as e.g. UV treatment, ozonation, and foam 578 579 fractionation to maintain a good water quality.

580 Compared to traditional aquaculture systems, it is possible to exert a higher degree of control on rearing 581 conditions in RAS and to collect and treat waste/effluents, reducing the impact on external environments. 582 Risk of escapees are reduced (and eliminated in fully closed systems), treatment of intake- and outlet 583 water improves disease control and reduces the need for medicine, it is possible to produce year-round, 584 and the most advanced RAS can be situated almost anywhere, including close to markets saving 585 transportation costs and ensuring full product traceability.

Recirculation technology has been used since 1980 to produce fry and fingerlings of different species, for instance for African catfish rearing in the Netherlands. Partial RAS technology has been applied in Chile since the 1990s as a means to comply with environmental regulations and save pumping energy for landbased production of exotic abalone (*Haliotis rufescens* and *Haliotis discuss hannai*) in seawater (Flores-Aguilar et al., 2007), and later for cultivation of endogenous species such as Chilean scallops (*Argopecten purpuratus*).

592

594 To produce fish for consumption, RAS are mainly used for finfish production, both indoor or outdoor. Semi-595 intensive, commercial RAS termed "Model-Trout-Farms" have been used in Denmark since 2004 as a

596 means to increase land-based production of rainbow trout (*Oncorhynchus mykiss*) while reducing the

- 597 impact on adjacent aquatic environments and complying to the EU Water Framework Directive (<u>Dalsgaard</u>
- 598 et al., 2013; EU-Water-Frame-Directive, 2000; Jokumsen and Svendsen, 2010). This concept, or parts of
- 599 it, has spread to other European countries although not as part of regulatory setups.

600 During the last decade, investments in RAS have intensified. These RAS are increasingly used in northern 601 Europe and Chile for producing Atlantic salmon smolt before transfer to sea cages (EUMOFA, 2020). 602 Growing smolt in RAS improves their growth and reduces subsequent sea-lice infestation problems in net 603 cages (<u>Clarke and Bostock, 2017</u>). Environmental concerns such as lack of freshwater and limited in-fjord 604 permits have further contributed to this development. An increasing number of intensive RAS have - or 605 are currently - being constructed globally aimed at producing primarily Atlantic salmon, but other species 606 are also targeted such as African catfish, barramundi, grouper, rainbow trout, seabass, seabream, 607 sturgeon, tilapia, pike perch, yellow tail kingfish (EUMOFA, 2020).

608 In Thailand, vertically integrated shrimp farming corporations developed RAS technology. These systems 609 are challenging when culturing crustaceans in water re-use systems as the molting cycles require extra 610 mineral inputs to compensate minerals lost with each shed exoskeleton. This requires the development of 611 special feeds to compensate for mineral deficiencies. In addition, cannibalism during molting leads to lower 612 survival, and sale prices cannot cover the high production costs in RAS. The price of shrimp broodstock is 613 very high, and broodstock do not molt frequently. Combined with high biosecurity and product quality, this 614 makes raising broodstock in RAS commercially feasible. To produce consumption size shrimp, however, 615 the production cost is too high. Some RAS farms to raise whiteleg shrimp or lobster have also been 616 constructed in Europe and elsewhere. These farms can be successful if producing for special markets that 617 fetch high prices.

Simple RAS technology has been applied in China since the 2000s for producing a range of species in both freshwater and seawater (shrimp, sturgeon, grouper, trout, turbot, sole, crab, salmon, seabass, pufferfish, sea cucumber, tilapia, a.o.). In the 2010s, RAS in China gained more governmental attention and advanced so-called "precision RAS" have been developed focusing on improving the engineering design, water treatment technology, and automation control (Huang, 2019).

On the downside, RAS are expensive to build and operate and they are challenging to manage requiring skilled workforce to ensure successful operation. They have a high carbon footprint in terms of compound feed (as applies to all compound fed aquaculture systems) and on-site energy requirements (especially for pumping) if electricity comes from fossil fuels (<u>Badiola et al., 2018; Martins et al., 2010; Midilli et al.,</u> 2012; <u>Samuel-Fitwi et al., 2013; Song et al., 2019; Van Rijn, 2013; Wilfart et al., 2013</u>). In addition, saltwater RAS require equipment that can withstand corrosion and risks of gas accumulation such as toxic H₂S and CO₂ are higher than in freshwater RAS (<u>EUMOFA, 2020</u>).

630 2.2.6 Culture-based fisheries, integrated multi-trophic aquaculture and aquaponic631 systems

632 <u>Culture-based fisheries</u>

633 Numerous seasonal water bodies are scattered throughout the world. Through culture-based fisheries, the 634 fish production from these water bodies can be enhanced, contributing to food security and enhancing rural 635 livelihoods (Subasinghe et al., 2013). Conversely, wetlands and deltas worldwide have been transformed 636 into agricultural or urban land, and sustaining the natural productivity and species diversity can be 637 challenging. Due to construction of dams and diversion of the natural water flow, fisheries production 638 declined, although traditionally, fisheries significantly contributed to income and food security in wetland 639 areas. In rice field areas in Cambodia, different forms of culture based fisheries, ranging from pure fisheries 640 to nearly fully controlled aquaculture are practiced side by side. Through proper management, these 641 systems contribute to food and nutrition security, rural livelihood diversification and income improvement, 642 and biodiversity conservation (Freed et al., 2020a; Freed et al., 2020b).

643 Considering the large number of existing water bodies suitable for culture-based fisheries, the potential 644 contribution to global fish production is large. <u>De Silva (2016)</u> calculated that if 20% of the small water 645 bodies in Asia would be used for culture-based fisheries, aiming for a production of 900 kg/ha, in Asia 646 alone 10.7 Mt could be produced annually. When developing culture-based fisheries, care should be taken 647 to produce within the capacity of the water bodies to neutralize the wastes resulting from feeding.

648 Integrated multi-trophic aquaculture

649 One concern of marine fed aquaculture is the release of organic and inorganic wastes (Schneider et al., 650 2005; Wang et al., 2012). In integrated multi-trophic aquaculture systems (Chopin et al., 2001; Neori et 651 al., 2007; Troell et al., 2009) fed species are linked to extractive species so that feed waste becomes food 652 for extractive species (Chopin, 2010; Chopin et al., 2001; Neori et al., 2007; Troell et al., 2003). The idea 653 behind the IMTA approach is that re-cycling of waste nutrients results in a reduced nutrient release into 654 the environment, while the overall productivity of the production system improves (Chopin et al., 2012). 655 Schneider et al. (2005) and Troell et al. (2003) reported variations in nutrient retention efficiency in IMTA 656 ranging between 2 to 100% depending on species, waste type, culture technique, and culture intensity. 657 Most IMTA systems are deployed at sea, with land-based IMTA reaching higher nutrient retention 658 efficiencies than open-water IMTA (Reid et al., 2017). Combining different species is not always easy, due 659 to differences in growth rate and seasonality and the large areas needed (Broch et al., 2013). Also the 660 retention efficiency of fish wastes by bivalves in IMTA is limited, because they need a minimum of 15-30% organic matter from fish waste in their diet to contribute to bioremediation (Cranford et al., 2013). 661 Consequently, IMTA at sea for bioremediation is still in its pilot phase, with the exception of China. Hughes, 662 Black (2016) suggested that extractive species can also obtain nutrients not originating from fed species. 663 664 This agrees with practices in China, where nitrogen inflow from land and phosphorous influx from open sea 665 are the major sources of nutrients in IMTA, with fed cage culture contributing a minor fraction of the 666 nutrient flow through the IMTA (Li et al., 2016; Mahmood et al., 2016a). Successful IMTA requires 667 significant architecture. Variations on this system have become routine in China including combinations of 668 seaweed, bivalves, finfish, sea cucumbers, and others (Liu Hui, Pers. Comm.). IMTA is theoretically 669 appealing in terms of sustainability, but from a management perspective, it is often not possible to optimize 670 production and marketing of multiple crops that are linked in these systems.

671 <u>Aquaponics</u>

672 In aquaponics, effluents from recirculating aquaculture systems (RAS) supply bioavailable nutrients for 673 plants (Paudel, 2020; Wongkiew et al., 2017). These are expensive systems (Palm et al., 2018) because 674 both a RAS and a hydroponic system need to be installed. Plants that do well in hydroponics also do well 675 in an aquaponic system. If nutrients for plants are lacking in the RAS effluent, extra fertilizers can be 676 applied to the plants (Eck et al., 2019; Maucieri et al., 2019). Whereas in coupled aquaponic systems, the 677 same water flow passes through all components of the aquaponic system, in a decoupled system water 678 flows through fish tanks, filters and hydroponics can be adjusted (Goddek et al., 2016; Monsees et al., 679 2017). Aquaponic systems are promising, because nutrient losses from aquaculture are reduced. A recent development are flocponic systems in which instead of a RAS a biofloc system is linked to a hydroponic 680 681 system (Pinho et al., 2017). Major constraints to profitability of aquaponic or flocponic systems include the 682 need to have access to high-value markets for vegetables and for fish, the broad range of know-how 683 involved, and the leading role of vegetables in securing financial success (Bosma et al., 2017). This in part 684 explains slow uptake by the industry.

685

686

688 3 Current issues and challenges in aquaculture

689 3.1 Environmental impact from aquaculture

690 3.1.1 Fed species

691 Concerns about the environmental impact of finfish aquaculture were flagged decades ago (Klinger and 692 Navlor, 2012; Navlor et al., 2000). Although considerable improvements were achieved in technologies to 693 reduce pollution through water treatment processes and better feeding methods (Xiao et al., 2019; Zhou 694 et al., 2018), the environmental impacts of fed aquaculture remains high. Globally, the demand for animal 695 products, including aquaculture species, will continue to grow due to population growth and rising incomes during the next decades (Hilborn et al., 2018). With fed aquaculture, impacts are mainly due to nutrients 696 697 provided in the feed that are not retained by the cultured species (Hilborn et al., 2018), and increased 698 energy inputs per unit surface area in relation to intensification (Ghamkhar et al., 2021). In a life cycle 699 assessment of aquaculture systems with different levels of culture intensity in Bangladesh, Henriksson et 700 al. (2018) found that across systems, fish yield is positively correlated with eutrophication and negatively correlated with water use, while no significant correlation was found with land use and climate change (CO2 701 702 emission). The feed conversion ratio (FCR) is an easy to measure indicator of nutrient use efficiency (NUE) 703 (Boyd et al., 2007), and is a better indicator of the environmental impact from fed aquaculture than the 704 type of farming system, the latter being either low- or high-intensity ponds, marine or freshwater cages, 705 or recirculating aquaculture systems (RAS) (Ghamkhar et al., 2021). Henriksson et al. (2018) identified 706 improving the NUE in aquaculture as the most effective way to reduce the environmental impacts from 707 aquaculture. This notion concurs with the concept of ecological intensification which, in addition to 708 improving the NUE of aquaculture, also aims to maintain or restore ecosystem functioning and diversity 709 (Aubin et al., 2017). Nevertheless, further improvements are necessary. For instance, in China, the total 710 nitrogen and phosphorous discharge through surface waters was similar for livestock and aquaculture, although the livestock sector is four times larger than the aquaculture sector (Zhang et al., 2015). Priority 711 712 should be given to recycling of nutrients as fertilizer, energy sources, or nutrient input for other organisms 713 (Chopin et al., 2012; Drózdz et al., 2020; Nhut et al., 2019; Wang et al., 2012).

714 Aquaculture in ponds has been intensified several times above the natural carrying capacity of a 715 conventional pond by providing intensive aeration (Itano et al., 2019; Jayanthi et al., 2021; Kumar et al., 716 2013) and removing waste from the system (Nhut et al., 2019). This helps to raise the production, but the 717 cost on the environment remains critical. Area-based waste water treatment in combination with renewable 718 energy generation and use can transform pollution mitigation from ponds into a productive and green 719 practice. Aquatic plants and sediments can trap waste nutrients while photovoltaic cells or wind energy 720 can reduce fossil fuel use. The economic feasibility of such interventions is location dependent. Integrating this approach for a cluster of farmers might be more feasible than for individual farmers. 721

722 3.1.2 Extractive systems

723 There is clearly a consensus that extractive species (filter-feeders; algae) exhibit orders of magnitude 724 lower ecological footprints than cultured fed species (e.g. tuna, salmon, shrimp). It is important, however, 725 to realize that if the aquaculture production does not properly consider aspects such as scale, site selection 726 and the effect of culture species on their environment, extractive species can also trigger environmental 727 issues. In the case of seaweed, the competition for space with seagrass meadows, addition of fertilizers, 728 and use of chemicals for controlling grazers, are potential risks that should be avoided. For example, in 729 South Korea and Japan, fertilization of seaweed farms has been banned (Hurd et al., 2014). Hence, careful 730 consideration of possible negative impacts on the coastal or offshore marine environment is needed.

- 731
- 732
- 733

Coastal pollution remains a significant challenge to shellfish aquaculture. The input of sediment, soils, and nutrients from numerous point and non-point sources affects growth and survival of the animals as well as their ability to be sold for human consumption. In addition to point sources, runoff from increasing amounts of impervious surfaces in many coastal areas can lead to waters being banned for use in shellfish cultivation.

739 Harmful algal blooms (HAB) have increased in frequency and range, and increased their seasonal growth 740 windows (Hallegraeff, 2010). These HAB occur regularly in areas occupied by shellfish aquaculture and 741 can have significant impacts on shellfish culture through mortalities or delayed harvests until toxins are 742 depurated (Basti et al., 2018; Matsuyama and Shumway, 2009; Shumway, 1990; Trainer et al., 2020). 743 Shellfish culturists are already addressing many climate-related issues and identifying new species for 744 culture, as well as implementing IMTA practices and HAB mitigation strategies. Climate change and HAB 745 interactions will be on-going and become increasingly important considerations for shellfish culturists. 746 Mitigation and remediation measures, coupled with the ability of bivalve molluscs to adapt to environmental 747 perturbations, will allow for the continued success of aquaculture systems.

748 Seaweed aquaculture systems are lagging behind in relation to other food sectors in breeding, pathogen 749 management, and adaptation of production systems to local nutrient, light and temperature conditions 750 (Buschmann et al., 2017; Navlor et al., 2021). Here too, disease interferes with farming efficiency. 751 Bacterial and viral outbreaks are especially high in intensively farmed seaweed systems, where disease 752 management can account for up to 50% of farm-variable costs (Barbier et al., 2019). New seaweed 753 cultivars with higher yield potential, disease resistance, nutritional qualities, and consumer attributes are 754 needed to ensure production, growth, and increased value for the industry, but an understanding of factors 755 that impact breeding programs and how they can affect wild seaweed populations is needed (Valero et al., 756 2017). In addition, the use of thalli fragmentation propagation techniques seems to have reduced the 757 genetic diversity of species like Eucheuma/Kappaphyccus and Agarophyton chilense (Eggertsen and 758 Halling, 2021; Guillemin et al., 2008) by unforeseen selection that appears to influence productivity and 759 resistance to pathogens and epiphytes (Hurtado et al., 2019). Nevertheless, other selected genotypes did 760 not show evidence of decreased productivity or loss of resistance to epiphytes. (Usandizaga et al., 2020). 761 Clearly these aspects require further in-depth studies to avoid the loss of genetic diversity or the 762 introduction genetically distinct individuals of a species in a region, thus introducing new genes that might 763 modify the adaptation capacity of the species under cultivation (Valero et al., 2017).

764 3.2 Land area

765 With fed aquaculture, ca. 0.1 - 0.4 ha of land area is needed to provide dietary plant ingredients to produce 766 1 MT of carnivorous and herbivorous/omnivorous animals, respectively (Aas et al., 2019; Boyd et al., 767 2007). An advantage of fish culture is that, in contrast to terrestrial animals, it is possible to move 768 production to aquatic environments (Boyd et al., 2020), reducing space and freshwater constraints. With 769 intensification, more fish are produced per unit area, but, because feed is the main nutrient input, 770 intensification has a minor impact on land area requirements per ton of fish produced. Compared to 771 terrestrial animal production, aquaculture requires less land than terrestrial animals, using only ca. 4% of 772 the total land area for crops dedicated to animal feed production (Hua et al., 2019; Troell et al., 2014b). 773 If the production volume of terrestrial animals from today onwards did not increase, with aquaculture 774 fulfilling the increased demand for animal protein until 2050, then only 10% extra cropland for feed 775 production would be needed for aquaculture feeds. Such a shift toward a human diet richer in aquaculture 776 products would save 70 to 76 million ha of cropland, an area close to the land mass of Turkey, compared 777 to a scenario with no shift in the balance between terrestrial and aquatic meat consumption (Froehlich et 778 <u>al., 2018</u>).

In shrimp farming, farmers and investors might abandon shrimp production sites after a few consecutive
crop failures due to disease. The area of these derelict ponds is large, covering an estimated few 100 000
ha globally, although exact data are missing. Reuse of these areas is possible, mainly by focusing on soil

and water quality in the wider production area.

Once corrective measures have been taken, shrimp production can recover, raising production and creating
 direct and indirect employment (Source: <u>http://kominfo.jatimprov.go.id/read/umum/38094</u>).

785 3.3 Water use

Compared to plant and terrestrial animal production, water use in aquaculture is relatively small and not well accounted for in food-water budgets (<u>Gephart et al., 2016</u>). Feed (ingredients, transport, processing), on-farm water use and fish processing are major categories of water use in aquaculture (<u>Gephart et al.,</u> 2016; <u>Henriksson et al., 2017</u>; <u>Verdegem and Bosma, 2009</u>). Intensification reduces the water use per unit production (<u>Henriksson et al., 2018</u>; <u>Verdegem et al., 2006</u>), making aquaculture more water efficient than meat production. With the expected growth in marine aquaculture (<u>Costello et al., 2020</u>; <u>Naylor et</u> <u>al., 2009</u>), the overall freshwater use efficiency by aquaculture will improve further.

793 3.4 Climate change and greenhouse gas emissions

794 The high diversity in species, culture systems, feeds, and salinity tolerance contributes to the resilience of 795 aquaculture to climate change, but does not make aquaculture disaster proof (Troell et al., 2014a). 796 Extreme weather events and sea level rise might cause flooding, water shortage, and changes in salinity, 797 temperature, and dissolved oxygen availability. Diseases might become more frequent, resulting in higher 798 losses and reduced animal welfare, and affecting the supply of external feed inputs to aquaculture (Reid 799 et al., 2019). Conversely, aquaculture contributes to global warming, due to increased energy and feed 800 inputs linked to intensification (Bostock et al., 2010) and greenhouse gas (GHG) emissions. Although GHG 801 emissions from aquaculture are modest compared to beef production (MacLeod et al., 2020), in semi-802 intensive coastal shrimp ponds or rice-fish systems methane emission is a concern, while there is nitrous 803 oxide (N₂O) volatilization in more intensive systems (<u>Henriksson et al., 2018</u>; <u>Yang et al., 2020</u>). Estimates 804 are that by 2030, aquaculture will release 5.7% of the anthropogenic N₂O emission. Culture systems that 805 trap nitrogen waste in biomass (microalgae, biofloc, seaweed, vegetables) are realistic options to reduce 806 N₂O emission from aquaculture (<u>Hu et al., 2012</u>).

Climate change will provide challenges for shellfish and seaweed production (Allison et al., 2011; FAO, 807 808 2016). As sea level rises, erosion of coastal areas increases, and the soils are washed into estuaries where 809 they increase turbidity and can affect the growth of submerged aquatic vegetation as well as disrupt 810 shellfish feeding. Increased temperatures, changes in sea level, precipitation, salinity, water circulation patterns, storm frequency and severity, and other factors will challenge shellfish farmers in different ways 811 812 in different geographic regions. It will be important to consider actual technological innovations in 813 aquaculture practices in response to climate change including material, procedural and informational 814 dimensions of practice (Lebel et al., 2020). It will be crucial to understand impacts of climate change at 815 the local level to promote sustainable shellfish culture. Global analyses for finfish species indicate regions 816 where culture condition improve and other regions where conditions will decline (Klinger et al., 2017; Oyinlola et al., 2018). Insights in local impacts are needed to inform policy and management officials. 817

818 Due to seawater level rise in some estuaries In South Sumatra, Indonesia, dikes of shrimp ponds had to 819 be raised 10 to 15 centimeters to protect against flooding. One mitigation measure to reduce economic 820 loss from extreme events due to climate change could be insurance. Although aquaculture insurance would 821 redeem the costs due to production loss or infrastructure damage, it will not bring back the lost production, 822 and still will reduce food security. However, the majority of producers across the aquaculture industry do 823 not have a 'crop' insurance. Large scale companies sometimes opt to not insure their crop (product), 824 assuming that the biosecurity and management systems employed are robust enough to guarantee 825 production, but insure their production infrastructure.

826 In the future, for small and medium sized farms, crop insurance may become more important. The technical 827 and physical infrastructure and management demanded by insurance companies may not be easy for 828 farmers to adopt Either way, the technical and management conditions could be assured by a specialized 829 technical service body, to the benefit of both parties.

830 3.5 Feed additives

High feed loads in highly and super-intensive culture systems quickly reduce water quality, stressing the immune system and the health of farm animals. A range of ingredients are used to enhance the immunocompetence and growth of the culture stock and/or improve water quality, including prebiotics, probiotics, enzymes, nucleotides, bacterial peptidoglycans, and carotenoids (<u>Boyd et al., 2020</u>). Most additives are mixed with, or coated on, the pelleted feed, but probiotics are often applied directly to the pond.

There are numerous manufacturers of probiotics, the effectiveness of which is not well defined. In addition, the action of a probiotic differs between fish species. More studies that test in depth the response of one fish species to one probiotic are needed, to get a better understanding of the effectiveness of the probiotic (<u>Ninawe and Selvin, 2009</u>). The numerous genome-sequencing and metabolomic tools can further help to elucidate how the immune system, nutrition, and growth affect nutrition (<u>Kumar et al., 2016</u>). Little is known of the long-term effects of probiotic use on the culture environment and needs further investigation.

843 3.6 Food security and poverty alleviation

844 Off the 20.5 million people employed full-time or part-time in aquaculture, most are small-scale workers, 845 of which a large fraction is engaged in other agricultural activities or off-farm employment. Others are 846 commercial producers, some of them employing workers during busy periods (Belton et al., 2018). With 847 future increasing incomes, the contribution of farmers engaged part-time in aquaculture to global 848 production will continue to decline (Edwards, 2015), while the contribution of commercial family-owned 849 producers will increase. The shift towards more intensive commercial production coincides with stocking 850 fewer species, favoring those species with a high market demand (Edwards, 2015). This might result in 851 less diversified and nutritious diets (Castine et al., 2017), but also improves food security of the poor 852 (Belton et al., 2020). To help small-scale aquaculture farmers to integrate into the value chain, education, 853 reliable property rights, credit, institutional support, and active engagement in decision making are highly 854 important (Salazar et al., 2018).

855 3.7 Technological innovation

856 3.7.1 Fed systems

857 Technological innovation should allow the aquaculture sector to upgrade productivity, lower costs, and 858 reduce environmental impacts. From a system perspective, key areas for innovation include water quality 859 maintenance, water purification (Xiao et al., 2019), feeding systems (Zhou et al., 2018), on-line 860 monitoring, and early warning systems (FAO, 2020; Hassan et al., 2016), and to share information bi-861 directionally through internet (Boyd et al., 2020; Gui et al., 2018; Hassan et al., 2016; Zhou et al., 2018). 862 The latter is only meaningful if supported with local education and capacity building (Lebel et al., 2020; 863 Weitzman, 2019), and guidance through best management practices (BMP). New technologies must be 864 communicated with farmers and integrated in sound BMP, in which attention to the use of chemicals and antibiotics, habitat destruction, fish escapees, and waste management is integrated (Boyd et al., 2007; 865 866 Diana et al., 2013). Measures can also be taken to reduce the effect of salt water intrusion in coastal zones due to climate change through land use planning and shifts in the selection of species adapted to the locally 867 868 prevailing salinity (<u>Nhung et al., 2019</u>).

- 870
- 871
- 872
- 873

⁸⁶⁹

874 Water purification technology and waste treatment in aquaculture got a boost with the increased interest 875 in RAS technology during the last decade, and the technology is available for any aquaculture operation to 876 treat wastes. Numerous farmers already apply aeration, concentrate and remove wastes from the culture 877 unit, or partially control toxic nitrogen levels. These are simple technologies, that if planned will raise farm 878 production in a cost effective way. A prerequisite is to have electricity next to the production facility, which 879 is more and more the case in rural areas around the world, and making intensification possible as described 880 in Chapter 2.3.2. These technological innovations are a necessity for farmers who wish to intensify, while 881 maintaining good water quality conducive to a healthy culture environment and production. In addition, it 882 creates options to collect and treat waste on farm (Chapter 4.3.2).

883 Installing a full RAS is different. At present, there are financial, operational, social, and marketing risks 884 associated with the development of intensive RAS (Figure 4). Investment and operational costs are 885 generally high when it comes to establishing a RAS. Most development is therefore seen in large-scale 886 systems assuming that "bigger is better" in terms of reducing capital expenditures/kg. Capital expenditures 887 are required upfront, time between initial investments and (potential) revenue is typically long, and many 888 things can go wrong before a system generates profit. Operating costs can also be higher than in more traditional farming due to expenditures on commodities such as 889 oxygen, CO₂ stripping, and electricity(https://www.undercurrentnews.com/2020/03/06/aquamaof-says-large-scale-salmon-ras-890

- 891 projects-will-produce-at-costs-of-around-3-kg/).
- 892 In addition, there are many legislative and marked related risks and challenges associated with RAS, and
- risk of failure should not be overlooked (<u>https://www.intrafish.com/finance/analysis-heres-a-list-of-high-</u>
- 894 profile-land-based-aquaculture-failures/2-1-712748, https://thefishsite.com/articles/rabobank-why-the-
- 895 <u>tide-is-turning-in-favour-of-ras-production, EUMOFA, 2020</u>) including in smaller-scale systems (Clarke and
- 896 <u>Bostock, 2017</u>).

897



Figure 4:

Four sets of risks for RAS operations facing large-scale RAS producers and investors. Reproduced with permission from <u>www.thefishsite.com/articles/raboba</u> <u>nk-why-the-tide-is-turning-in-</u> <u>favour-of-ras-production</u>.

The risks facing RAS producers and their investors © Rabobank

898 While freshwater RAS has proven successful for producing fry and juveniles of several species including 899 salmon smolts, and for rearing portion-sized trout, saltwater RAS is still a challenge. Gear that can 900 withstand corrosion is required, biofilter performance may be hampered at higher salinity (<u>Kinyage et al.,</u> 901 <u>2019</u>), and even a minor accumulation of sludge can result in sudden formation of H₂S with devastating 902 consequences. Currently no sensors for reliable detection and early warning of H₂S are in use, and sensor 903 technology in general is a ubiquitous issue in both fresh and saltwater RAS.

System management and qualified staff are key to successful RAS operations (Bregnballe, 2015; EUMOFA,
 2020), in addition to good knowledge of cultured species biology. Staff need to be skilled in water quality
 monitoring and assessment, in operating different treatment technologies (including end-of-pipe
 solutions), and in managing surveillance and backup systems.

- 909 Market value of the cultivated species must justify higher production costs. Product value may change over
- 910 time due, e.g., to competition and changes in consumer preferences and willingness to pay extra for
- 911 seafood cultivated in RAS. In line with this, it is essential that product quality be optimized and that, for
- 912 example, an ubiquitous issue with off-flavour is reduced and preferably avoided (<u>EUMOFA, 2020</u>).
- 913 Site selection, permits, and licenses for establishing a RAS facility can be difficult, time consuming, and 914 costly, and may involve a high degree of stakeholder involvement. Getting a permit in the USA may, for 915 example, add up to 5% of the total budget (Ramboll Webinar on "Water management in RAS systems,
- 916 Maine & Denmark". November 19, 2020), and in Europe, processes of obtaining a permit are challenged
- by interactions between local, regional, national, and EU legislation (EUMOFA, 2020).

918 3.7.2 Extractive systems

- 919 Improved mitigation strategies for the control of biofouling of shellfish grow-out equipment are still needed.
 920 While there are numerous physical methods for removal of biofouling (Watson et al., 2009), they are labour
 921 intensive and can result in damaged crops. Promising new research and development of environmentally
 922 friendly coatings have shown good results (Shumway, in press) and will lead to reduced labour and increase
 923 yield per area, which in turn will lead to increased production of crops.
- 924 Improved methods to assess biomass during production in bottom culture will continue to be a challenge.
 925 Bottom culture methods are still quite primitive and utilize harvest equipment that has changed little during
 926 the past century. They lack the ability to assess biomass quickly and effectively, and improved techniques
- 927 will increase production and expand markets.
- 928 There is a need for advanced technology in production to increase output and lower labor input to make 929 businesses more profitable and competitive. Some of this technology can be adapted from agricultural 930 industries. Two recent examples include planting and harvest using technology adapted from the bulb 931 industry and sorting and grading technology that evolved from that developed for apples. A long-elusive 932 component of oyster processing has been automated shucking machinery that could remove meats from 933 the shells quickly without manual labor. Current advances in sensing and computer-aided machinery should make this possible. The economic and managerial feasibility of production systems will ultimately drive 934 their development and acceptance. Inclusion of increased automation will ultimately depend upon the 935 936 increased additional costs as compared to the additional economic benefits.
- 937 The majority of marine plants are grown on suspended long-lines or suspended nets. Some red algae are 938 harvested from the wild or grown in bottom culture in shallow water, but they represent a minor percentage 939 of the global production. Harvesting of seaweeds is labour intensive, and, here too, the industry will benefit 940 from wider use of existing or improved semi-automated harvesting techniques. Improving existing 941 technology to seed and deploy long-lines effectively, with minimum labour input is presently getting a lot 942 of attention, and will help to reduce labour inputs.

943 4 Priorities for future aquaculture development

944 4.1 Climate change

The relation between aquaculture and climate change has been recognized and studied for some time (Cochrane et al., 2009). During the last 20 years, the aquaculture industry improved from a technological, governance, management and siting perspective. This made aquaculture better able to cope with harmful algal blooms (Chapter 3.4), disease (Chapters 2.3.3 and 3.1.1) and extreme weather events (Chapter 3.4). However, generalization is difficult. There is a need to better monitor the effects of environmental changes on on-farm production to be able to validate concepts developed in laboratory simulations across production environments and culture species (Naylor et al., 2021).

953 Similarly, little information is available on GHG from aquaculture facilities (Chapter 3.4), because emissions
 954 are influenced by the culture environment and system management. There is insufficient evidence that
 955 study outcomes can be generalized, nor about which management practices minimize GHG emission from
 956 specific culture systems. Important parameters include the type of ingredients used in feed formulation,
 957 NUE of carbon and nitrogen, and the dissolved oxygen dynamics.

958 More research is needed to find ways to improve the energy use efficiency in fed aquaculture. One approach 959 is the nutritious food concept (Chapter 2.2.1), but much more research is needed. In addition, contributing 960 to circularity by making use of crop residues in aquaculture feeds contributes to reducing the energy 961 expenditure of food systems (Chapter 4.3.1).

962

963 4.2 Sustainable aquaculture

964 4.2.1 Improved pelleted diets and fish meal and fish oil use

A major public concern is the use of wild small pelagic species as forage fish to culture high value finfish 965 966 species (Diana et al., 2013; Little et al., 2016; Naylor et al., 2000). During the last decade, substantial 967 improvements have been achieved in reducing the percentage of fishmeal and fish oil in pelleted feeds 968 (Naylor et al., 2021). Improved insights in the requirements for some 40+ essential nutrients as opposed 969 to ingredients (e.g. fishmeal and fish oil) allow the industry to include a broader range of plant and animal 970 ingredients in fish feed. This includes re-using products like crop residues otherwise not used, making 971 aquaculture part of the circular economy (Van Zanten et al., 2019). Specific limiting essential nutrients, 972 such as amino acids, fatty acids, vitamins, minerals, and trace elements, can be added in the diet to fulfil 973 the nutrient requirements of the target species (Boyd et al., 2020). In addition, by enzyme or probiotic 974 inclusion in the diet, the utilization efficiency by the fish of low quality ingredients can be upgraded, 975 enlarging further the range of ingredients that can be used in fish diets (Maas et al., 2018; Maas et al., 976 2020; Maas et al., 2021). This combined approach gives the industry many options to reduce the use of 977 fishmeal and fish oil in fish diets and to test a broad range of alternative ingredients to fishmeal or fish oil. 978 Promising ingredients tested include fishery and aquaculture by-products, food waste, insects, microbial 979 biomass (e.g. microalgae, bacteria, yeasts), and macroalgae (Ghamkhar et al., 2021; Hua et al., 2019; 980 Matassa et al., 2016). Some ingredients are already better optimized for use in aquaculture than others. 981 For example, Maiolo et al. (2020) tested the effect of insect meal, poultry by-product meal, and microalgal 982 biomass as partial substitutes for fish meal, assessing the environmental effects through life cycle analysis 983 (LCA). Poultry by-product meal has the smallest environmental impact, showing optimal environmental 984 performance, while the environmental impact of insect meal and microalgal biomass could be further 985 reduced by improving the NUE and reducing energy use. For food wastes and microbial biomass products, 986 considering the economics of commercial production, supply and supply consistency, are still large hurdles 987 that must be overcome to be able to meet demand (<u>Hua et al., 2019</u>). This makes insects and microalgae 988 currently the most promising candidates for fishmeal and fish oil replacement, respectively, in fish diets 989 (Cottrell et al., 2020). Nevertheless, a wide range of factors need to be considered in concert (e.g. 990 substitutability, nutritional profiles, scalability, environmental impacts) when aiming for sustainable growth 991 in production. Doing so often leads to opposing conclusions, highlighting the need to integrate multi-992 objective analyses in the decision process (Pelletier et al., 2018).

A result of shifting from ingredient availability to essential nutrient requirements to guide diet formulation is that the differences in the trophic level of diets between herbivores, omnivore and carnivores reduce and converge (<u>Cottrell et al., 2021</u>), while resources that previously could not be used can now be recycled through fish diets. Once replacements for fishmeal and fish oil become available at a competitive price and in sufficient quantity, then large-scale production of carnivorous marine fish species in response to consumer demands might develop quickly (<u>Gephart et al., 2020</u>).

1000 One development is that strains developed for fast growth require a protein and nutrient-rich high quality 1001 diet to be able to maintain fast growth. This makes the feed expensive while feed companies aim to 1002 formulate diets at a cost that allow the farmer to make a profit. Farmers and consumers should have 1003 insight into the quality of the diet and the selection of ingredients used. This is politically and 1004 environmentally sensitive information, requiring full attention from the feed companies. (source: personal 1005 communication during interview discussion with feed manufacturing sector specialists from Australia, India, 1006 Indonesia, and Thailand. 2020-21). Conversely, not all farmers have to stock fast growing strains in 1007 intensive systems. For them there is a need to have access to strains that grow well on less expensive 1008 feeds containing recycled agricultural wastes.

1009

1010 4.2.2 Waste treatment technology and RAS development

1011 Today, numerous aquaculture farms use technologies first applied in RAS (Chapter 3.7.2). Having the 1012 option of temporarily recirculating the water e.g., by simple airlifts, may help a farmer circumvent 1013 upstream disease outbreaks or water shortages. This also reduces the use of antibiotics and therapeutants. 27

1014 Continuous removal of solids (fish faeces and uneaten feed) prior to discharge, e.g., via sludge cones, 1015 drum filters, swirl separators, or fix-bed biofilters, may reduce the discharge of particulate organic matter 1016 and phosphorus considerably, benefitting the recipient and improving internal water quality. As stocking density or recirculation intensity increases, biofiltration will be necessary to convert ammonia to nitrate. 1017 1018 Here, use of locally available filter material such as coconut shells rather than commercially manufactured 1019 plastic bio-elements might be both cheaper and more sustainable. Generally, use of locally manufactured 1020 equipment adapted to resist local conditions, as well as aquafeed developed using local ingredients, might 1021 ensure a higher rate of success.

1022 To reduce eutrophication further, nitrate may be removed from farm effluent by treating it in simple, low-1023 cost denitrifying bioreactors. These are water-logged beds filled with locally available, slow-releasing carbon sources such as woodchips or coconut husks (Rambags et al., 2019; von Ahnen et al., 2018). In 1024 1025 addition to removing nitrate, denitrifying bioreactors are known to be effective in reducing microbial 1026 contaminants, providing complimentary disinfection (Rambags et al., 2019). Water treated in denitrifying 1027 bioreactors may potentially be recycled back to ponds following e.g., sand filter filtration, to remove 1028 harmful or toxic substances potentially released from the filter material (Lepine et al., 2021; Lindholm-1029 Lehto et al., 2020).

- Finally, education and training are imperative if these technologies first developed for RAS are to contributemassively to sustainable production of healthy aquatic foods in the future.
- 1032

1033 4.2.3 Disease prevention and biosecurity

1034 Increasing demand for aquaculture products and the expansion of seafood stimulate intensification, and 1035 with intensification reliance on high density monocultures which are vulnerable to disease. Predictions are that in tropical shrimp production alone, 40% of the production will be lost annually to disease-related 1036 1037 mortality (Stentiford et al., 2012). Current understanding of how antibiotics in the farming environment 1038 influence disease occurrence is still limited. For instance, temperature has major impacts on disease 1039 susceptibility of shrimp, but insights into how to manipulate temperature or any other abiotic factor to control disease are still lacking (Millard et al., 2020). Knowledge on balances/imbalances in the microbiome 1040 1041 of aquaculture systems is still in its infancy, although quickly developing (Infante-Villamil et al., 2020). 1042 More research on creating a healthy environment for culture organisms is needed, requiring more meta-1043 analyses and cross-sectoral studies in addition to disciplinary research (Reverter et al., 2021). The latter 1044 is important in view of global warming and ocean acidification (Green et al., 2014; Rowley et al., 2014).

1046 Although the root of many disease problems in aquaculture lies to a high degree with the culture system 1047 applied, the main goal of the industry and the research community is to find effective disease treatments, 1048 which can be a time-consuming and long-term effort. For instance, in shrimp farming there are currently 1049 no effective treatments for most viral, but also for some bacterial diseases, even after decades of research 1050 (Flegel, 2019). Through development and application of best management practices (BMP), including 1051 biosecurity measures, the industry tries to minimize the impact of disease. The principal key to success is 1052 how well recommended practices of the BMP help to maintain or restore a healthy production environment 1053 for the farmed animals.

1054 In relation to biosecurity there is a high level of misunderstanding among stakeholders regarding "the 1055 fundamental principles and practices of aquaculture pond management". When aiming for disease control, 1056 the main objectives are (1) to prevent a pathogen enters the farm, and (2) to prevent any pathogens 1057 present from spreading around the farm. These objectives must be part of a quality assurance (QA) 1058 programme. Aquaculture farms are often clustered, therefore requiring coordination and cooperation 1059 among stakeholders in applying biosecurity. If all actors involved apply the biosecurity measures correctly, 1060 then disease incidences can be drastically reduced. Large industrial farms are more successful in applying 1061 biosecurity, as they are able to implement and cover the costs of strict quality assurance/quality control 1062 (QA/QC) protocols. Small-scale farmers need to work collectively to implement the biosecurity measures 1063 to be able to bear the QA/QC costs. Latt (2019) suggested protocols for a QC laboratory to inform the right 1064 QA actions for clusters of shrimp hatcheries, that can be collectively implemented by a cluster of 4-5 small or medium-sized hatcheries. Hence, effective biosecurity, including good QA/QC protocols can be 1065 1066 implemented. Successful biosecurity application will require adequate training of farmers in aquaculture 1067 pond management and enforcement of government regulations, conductive to creating and maintaining a 1068 healthy production environment. Existing loopholes breaching biosecurity enforcement should be 1069 prevented, requiring strong political will and multi-stakeholder cooperation, involving government, 1070 education and the aquaculture and food industry stakeholders.

1071 In shrimp farming, biosecurity is further enhanced by the provision of specific pathogen free (SPF), specific 1072 pathogen tolerant (SPT) or specific pathogen resistant (SPR) post larvae (PL), which today is common 1073 practice across the industry. This development concurs with efforts to reduce pond size, in combination 1074 with better control on the application of biosecurity protocols. Similar developments are observed in China, 1075 Vietnam, Thailand and Indonesia, all major players in the shrimp industry.

1076

1077 4.3 Monitoring and early warning systems

1078 Automation in any system, if applied judiciously, can improve production and profits. For example, the 1079 introduction and implementation of 'smart technology' and robotics to enhance shellfish farm production 1080 is currently being explored. Technologies such as artificial intelligence, Big Data, Internet-of-Things, and 1081 robotics are used routinely in modern agriculture and result in increased efficiency and lowered costs. 1082 These technologies are not yet readily applicable throughout the aquaculture industry, although they are 1083 applied in automated feeding technology or remote monitoring of weather and water quality developments 1084 around cage culture operations at sea. For shellfish culture, it is proposed that use of these technologies 1085 could improve bottom culture of oysters by aiding planting oysters accurately in the right densities. Site 1086 selection of finfish, shellfish, or seaweed operations at sea can be guided by means of GIS and farm-scale 1087 models (Ferreira et al., 2009; Silva et al., 2011). Advanced sensing gear will allow growers to determine 1088 growth and mortality of their crops for management decision, and the technology used to develop 1089 advanced sensing and imaging that will allow the ability to monitor crops in situ, reduce mortality, and 1090 lead to advanced and selective harvest methods (D. Webster, Personal Communication). More specifically, 1091 in shellfish farming, technological developments in precision aquaculture can be used to promote 1092 autonomous and continuous monitoring using underwater sensors that observe, analyze, interpret, and 1093 assist decision-making support for farm operations (Donncha and Grant, 2019).

1094 While there are still issues to be addressed including long-term maintenance of the sensors, limited battery 1095 capacity of the sensors, high energy consumption during signal transmission, environmental impacts on 1096 the sensors, and cost (<u>Parra et al., 2018</u>), these are promising technologies that will provide further means 1097 to improve the efficiency of shellfish aquaculture.

1098 Monitoring systems in source waters as well as disease surveys in the near environment have been part 1099 of planning and management of production in some vertically integrated aquaculture industries since early 1100 2000. Such programmes and practices require significant resources, both financial and human, to produce 1101 beneficial outcomes. Small and medium-sized enterprises should organize in clusters or farmer group, and 1102 closely cooperate and collaborate with government and academic sectors to develop and implement better 1103 planning and management of productions. Weather predictions and climate change impact scenario, have 1104 also been incorporated into production, marketing, and strategy formulation of certain medium- and large-1105 scale shrimp farming enterprises since late 2000. If properly implemented and acted upon immediately, 1106 mitigation of climate change-related impact on production is possible allowing the farmers, the principal 1107 actors, to react swiftly.

1108

1109 4.4 Enabling environment

1110 4.4.1 Institutions and regulation

Shellfish aquaculture has a bright future. It is a sustainable source of protein and can give a major boost to local economies and food security globally. Several species will likely continue to dominate the international shellfish aquaculture community, most notably clams, oysters, scallops, and mussels. In addition, other less recognized species will be successfully cultured and provide jobs on more local levels, e.g. cockles. Not all species, systems, and technologies will be commercially feasible, but without continued research, positive attitudes, and a willingness to explore and accept new possibilities, there can be no growth.

1118 The same holds for seaweed aquaculture. It is sustainable, contributes directly and indirectly to food 1119 systems, can mitigate environmental impacts, and generates income. The main species presently cultured 1120 will remain so in the near future.

To continue on a positive growth trajectory of extractive species, however, there will need to be improved 1121 1122 acceptance by the public and modification of rules, laws, and regulations that currently hamper 1123 establishment of new farms and expansion of existing ones. For example, for shellfish farming, 1124 government regulations have been a significant factor in limiting or preventing the establishment and 1125 expansion of shellfish aquaculture in many regions globally. Development of nearshore shellfish 1126 aquaculture is increasingly constrained by space, economics, human health, societal concerns, and 1127 environmental issues (Cheney et al., 2010). The same holds, to a lesser extent, for cage culture at sea 1128 (Chapter 2.7.1).

1129

1130 4.4.2 Education and training

1131 To raise public awareness and acceptance by the public, expanded educational efforts are needed.

Proactive programs on aquaculture should be created to educate communities about the benefits to the economy, employment, and food security. For extractive species, benefits to the environment should be stressed.

1135 The development of offshore aquaculture has raised a new level of concern. Offshore aquaculture will

1136 happen in countries and waters where the regulatory systems allow it to proceed. Federal and

1137 international agencies are involved in the permitting processes and often do not work collaboratively or

even proactively. Significant legislation will be required to enable development of those productive and

1139 lucrative offshore waters.

1140 As with most change, there will be a period of adjustment and acceptance by both industry and consumers, 1141 and focused education and communication programs can be a great asset. For aquaculture to continue 1142 expansion, a range of support programmes are needed. There needs to be strong and effective 1143 communication regarding delivering food security to local communities in the short term, while 1144 simultaneously educating them and encouraging partnerships working toward resource sustainability and 1145 community development in the longer term. These programs and their implementation will vary according 1146 to local needs, but should focus on school programs, youth programs, and public education. These programs should include general education regarding the benefits of extractive aquaculture to the 1147 environment and the community, but also should include programs to teach production skills to train future 1148 1149 aquaculture leaders.

1150 Education related to disease prevention and best management practices is important to enhance production 1151 stability in aquaculture. The education programme should be developed as simple as possible, addressing 1152 a wide audience with varying educational background. Reasoning should be realistic, logical, technically 1153 sound, and acceptable. Culture system, standard operational procedure, biosecurity, management regime, 1154 and sustainability principles should be included and enhance critical thinking through WH-questioning 1155 (what, when, where, who, whom, which, whose, why, and how) for better understanding. Preferentially, each programme should be designed with outcome-based implementation in mind. Delivery of the program 1156 1157 could be monitored and continuously improved with the program developer and trainer sharing 1158 accountability on the targeted outcome.

1159

1160 4.5 Aquaculture development priorities

1161 4.5.1 Contribution of aquaculture to the 2000 & 2010 development priorities

- Some of the strategies and recommendations given during the Global Conferences on Aquaculture held in2000 in Bangkok and in 2010 in Phuket in Thailand relate to aquaculture systems. These
- recommendations are copied below, and advances made since 2000 are briefly outlined.
- Aquaculture should adopt farming practices that ensure environmental sustainability, based on environmentally sound technologies and resource efficient farming systems.
- 1167Progress: In response to public pressure, the environmental sustainability of aquaculture farming1168practices improved, benefiting from technological advances and improved biological insights1169(Chapters 3.1 and 4.2.2). Public pressure will not decline and the sector will continue to improve1170its environmental performance during the next decade.
- Aquaculture should aim to increase its impact on rural development and poverty alleviation, making
 aquaculture an integral part of rural development programs.
- 1173Progress: Aquaculture contributed significantly to livelihoods of small-scale farmers, poverty1174alleviation, and food security. Overall, the sector has intensified, including small-scale farmers1175who benefitted from advances in rural infrastructure (accessibility, electricity), technology (feed1176technology, more and better farm equipment), and for some species improved breeds (Chapters11772.2.1 and 2.2.2). There is a need to provide training, so farmers can take full benefit from these1178technological advancements (Chapter 4.4.2).
- Develop a well-documented set of varied and adaptable technologies and systems allowing producers to make the best use of their local environment, and to make well informed decisions on the production system and species they will use. This also includes making aquaculture systems more resilient to successfully face the uncertainties and risks wrought by climate change.
- 1183Progress: Aquaculture systems management has improved, resulting in less frequent production1184failures. Farm management procedures have been developed to make farming systems less1185vulnerable to disease-related production losses when implemented correctly. This is an ongoing1186effort, requiring more focus on training during the next decade (Chapter 3.1.1).
- Pay attention to culture-based fisheries, sustainable stock enhancement, and ranching programs
 aiming to make efficient use of under-utilized, new or degraded resources.

- 1189Progress: Culture-based fisheries can make a significant contribution to global aquaculture1190production, and can be integrated in agricultural areas (Chapter 2.2.6). Culture-based fisheries1191practices are location-dependent and guidelines are missing as to how to develop sustainable1192culture-based fisheries practices adaptable to local contexts. This is a priority for future1193development.
- Put emphasis on integrated aquaculture systems to improve environmental performance of
 aquaculture.
- 1196Progress: Numerous studies have investigated integrated aquaculture systems and documented1197improved nutrient use efficiency and environmental performance. The high knowledge1198requirements, however, and difficulties to make these systems economically viable hamper1199adoption. Only in various coastal bays in China are integrated aquaculture systems successful1200(Chapter 2.2.6). Integrated culture of fish and shrimp helps to reduce disease occurrence in1201shrimp farming (Chapter 2.2.2), and should be promoted.
- Further develop new technologies including recirculating aquaculture, offshore cage culture, artificial
 upwelling and ecosystem-based food web management.
- 1204Progress: Recirculating aquaculture systems have strongly developed since 2000 (Chapter 2.2.5)1205and technologies are spreading to outdoor systems, both for finfish and crustaceans (Chapters12063.7.1 and 4.2.2). Both high-intensity RAS and offshore aquaculture are costly, and, at present1207only feasible for expensive species. Ecosystem based management should receive more1208attention: managing the food web through feed formulation raises the nutrient use efficiency in1209pond systems and reduces the environmental impact (Chapter 2.2.1), but more research is1210needed.
- Make aquaculture resilient to successfully face the uncertainties and risks wrought by climate change
 Progress: Aquaculture systems have become more resilient, but more research is needed to gain
 better insight as to how different species, production environments, and management practices
 affect resilience to climate change (Chapter 4.1).
- Accelerate the untapped potential for aquaculture development outside Asia.
- 1216Progress: High growth of aquaculture development in Africa and Latin America is recognized and1217is driven by fast growth in a limited number of countries (Chapter 2.1). Stimulating aquaculture1218development in more countries on each continent will help to accelerate aquaculture1219development outside Asia (Chapter 4.5.2).
- 1220

1221 4.5.2 Priorities until 2030

1222 Aquaculture production, which is presently growing (Chapter 2), will continue to do so in the nearby future 1223 (Garlock et al., 2020; Little et al., 2016). This will predictably lead to ~18% higher world aquatic food 1224 consumption (including capture fisheries) in 2030 compared to 2018 (FAO, 2020). The successful 1225 integration of aquaculture into the global food system since 1997 (Naylor et al., 2021) has contributed to 1226 the provision of sufficient calories from aquatic foods. Healthy food is, however, not only about cheap 1227 calories, but also about foods that contribute to a nutritious diet, the latter providing a balanced mix of 1228 bioaccessible nutrients to sustain a healthy and active lifestyle. Aquatic foods play an important role in 1229 providing such a healthy nutritious diet (Thilsted et al., 2016). The latter requires a 'nutrition-sensitive' 1230 food system which provides a diverse and nutritional complete set of foods, is embedded in society and contributes to sustainable livelihoods (Uccello et al., 2017). Aquaculture can contribute to nutrition-1231 1232 sensitive food systems that alleviate health inequities under various scenarios ranging between endless 1233 growth versus staying within planetary boundaries and between regionalized versus globalized food 1234 systems (Figure 5) (Gephart et al., 2020).



Figure 5

Scenarios for future growth of aquaculture considering endless growth versus sustainable growth (doughnut economics), focusing on globalization versus regional development (<u>Gephart et</u> al., 2020).

32

1235

Today the world produces enough food to meet the human nutritional needs of nearly 10 billion people in 2050, provided all people get assured access to food and human feeding habits change. The latter includes reducing consumption of animal foods and replacing them with plant-based alternatives. This includes direct human consumption of crops such as maize, now fed to animals (Berners-Lee et al., 2018) (Figure 6).



1241

1242 Most people prefer not to forsake animal food as suggested by Berners-Lee et al. (2018) considering fed 1243 aquaculture a legitimate and sustainable way of producing healthy food, and seeing room for further growth 1244 of aquaculture during the next decade. Important questions are (1) how much should aquaculture 1245 production expand during the next decade, and which products should be best produced, where, and how? 1246 Answers should consider the urgent challenges to the global food system, including aquaculture, to address 1247 global warming and to produce food within planetary boundaries. One possible solution is to develop a 1248 doughnut economy (Raworth, 2017) producing products so that nobody in society is left behind (inner ring 1249 of the doughnut) and leaving a healthy planet behind for generations to come (outer ring of the doughnut).

1250 Sustainable, nutrition-sensitive food systems, when the resource base permits, are possible under each of 1251 the scenarios presented in Figure 5, but within a doughnut economy, only the scenarios of 'food 1252 sovereignty' or 'blue internationalism' apply. The first scenario focuses on local production by smallholders, 1253 the second on international trade. Both scenarios focus on sustainable production within the local context. 1254 Considering aquaculture, sustainable production within the local context would be easier if aquaculture was 1255 spread more evenly across the world. Especially in Africa and Latin America, there is room for aquaculture 1256 development with opportunities to make aquaculture an integral component of nutrition-sensitive food 1257 systems (Chapter 2.2.).

1259 Considering the above, criteria to judge aquaculture development during the next decade should not focus 1260 primarily on global growth, except for extractive species. The industry found ways to improve its 1261 environmental performance during the last decades, and as societal pressure to improve upon 1262 environmental performance will increase further, the aquaculture sector will continue to improve its 1263 environmental performance. Therefore, the main focus should be:

1264 (1) to develop sustainable aquaculture systems imbedded within local food systems, with focus on 1265 a. minimizing nutrient losses from aquaculture systems. This relates to solid, dissolved, and 1266 gaseous (including GHGs) waste. Special attention should be given to development of innovative low-cost effective ways to reduce, collect, or process wastes from aquaculture 1267 1268 systems. b. maximizing nutrient utilization efficiency of harvested products, with special attention to 1269 1270 polycultures and multi-trophic systems, 1271 c. recycling nutrients across agriculture - aquaculture farming systems contributing to 1272 circularity, 1273 d. water re-use, minimizing water use in freshwater and brackish water aquaculture. 1274 (2) to embed aquaculture within nutrition-sensitive food systems. This requires monitoring the 1275 nutritional value from aquaculture products harvested from different aquaculture systems. 1276 (3) to provide equitable income for stakeholders, including farming households, along the aquaculture 1277 value chain. This requires multi-objective analysis, including tradeoffs between production, 1278 income, nutrient use efficiency, environment, and nature. 1279 (4) to increase the contribution from molluscan culture to global aquaculture, with focus on 1280 a. developing simple and logical procedures, laws and regulations guiding the development 1281 of molluscan farming. b. education campaigns to inform the public and farmers on the benefits and advantages of 1282 1283 molluscan farming in areas suited for molluscan culture. 1284 stimulating the development and implementation of 'smart technology' and robotics to c. 1285 enhance especially shellfish production at sea. (5) to stimulate further growth of seaweed aquaculture not only in Asia, but also on other continents. 1286 1287 Special attention should be given to: 1288 a. Selecting culture species adapted to the local climate and environment, 1289 b. qualifying and quantifying ecosystem services provided by aquatic plants, 1290 c. implementing smart technology and robotics to enhance seaweed culture. (6) to stimulate aquaculture development outside Asia, especially in Africa and Latin America. 1291 1292 (7) to address climate change, including 1293 documentation of the effects of physical and chemical changes in the production a. 1294 environment on production, animal health and resilience for a wide array of culture 1295 systems and species; 1296 b. develop early warning systems, adapted to local contexts, to insulate 1297 farmers/communities against catastrophic effects.

1298

1299 5 References

- Aas, T.S., Ytrestøyl, T., Åsgård, T., 2019. Utilization of feed resources in the production of Atlantic salmon
 (Salmo salar) in Norway: An update for 2016. Aquaculture Reports. 15.
- 1302Ahmed, M., Lorica, M.H., 2002. Improving developing country food security through aquaculture development -1303Lessons from Asia. Food Policy. 27, 125-141.
- 1304Ahmed, N., Ward, J.D., Saint, C.P., 2014. Can integrated aquaculture-agriculture (IAA) produce "more crop per1305drop"? Food Security. 6, 767-779.
- Alday-Sanz, V., 2018. Specific pathogen free (SPF), specific pathogen resistant (SPR) and specific pathogen
 tolerant (SPT) as part of the biosecurity strategy for whiteleg shrimp (Penaeus vannamei boone 1931).
 Asian Fisheries Science. 31, 112-120.
- 1309Allison, E.H., Badjeck, M., Meinhold, K., 2011. The Implications of Global Climate Change for Molluscan1310Aquaculture, Shellfish Aquaculture and the Environment, pp. 461-490.
- 1311Asaduzzaman, M., Wahab, M.A., Verdegem, M.C.J., Benerjee, S., Akter, T., Hasan, M.M., Azim, M.E., 2009.1312Effects of addition of tilapia Oreochromis niloticus and substrates for periphyton developments on pond1313ecology and production in C/N-controlled freshwater prawn Macrobrachium rosenbergii farming1314systems. Aquaculture. 287, 371-380.
- Aubin, J., Callier, M., Rey-Valette, H., Mathé, S., Wilfart, A., Legendre, M., Slembrouck, J., Caruso, D., Chia, E., Masson, G., Blancheton, J.P., Ediwarman, Haryadi, J., Prihadi, T.H., de Matos Casaca, J., Tamassia, S.T.J., Tocqueville, A., Fontaine, P., 2017. Implementing ecological intensification in fish farming: definition and principles from contrasting experiences. Reviews in Aquaculture. 11, 149-167.
- Avnimelech, Y., 2007. Feeding with microbial flocs by tilapia in minimal discharge bio-flocs technology ponds.
 Aquaculture. 264, 140-147.
- 1321Badiola, M., Basurko, O., Piedrahita, R., Hundley, P., Mendiola, D., 2018. Energy use in recirculating1322aquaculture systems (RAS): a review. Aquacultural engineering. 81, 57-70.
- Barbier, M., Charrier, B., Araujo, R., Holdt, S.L., Jacquemin, B., Rebours, C., 2019. PEGASUS PHYCOMORPH
 European guidelines for a sustainable aquaculture of seaweeds. in: Barbier, M., Charrier, B. (Eds.),
 COST action FA1406, Roscoff, France.
- Basti, L., Hégaret, H., Shumway, S.E., 2018. Harmful algal blooms and shellfish. Harmful Algal Blooms: A
 Compendium Desk Reference; John Wiley & Sons, Ltd.: Hoboken, NJ, USA, 135-190.
- 1328Belton, B., Bush, S.R., Little, D.C., 2018. Not just for the wealthy: Rethinking farmed fish consumption in the1329Global South. Global Food Security. 16, 85-92.
- Belton, B., Little, D.C., Zhang, W., Edwards, P., Skladany, M., Thilsted, S.H., 2020. Farming fish in the sea will
 not nourish the world. Nature Communications. 11.
- 1332Berners-Lee, M., Kennelly, C., Watson, R., Hewitt, C.N., 2018. Current global food production is sufficient to1333meet human nutritional needs in 2050 provided there is radical societal adaptation. Elementa-Science1334of the Anthropocene. 6.
- 1335Bosma, R.H., Lacambra, L., Landstra, Y., Perini, C., Poulie, J., Schwaner, M.J., Yin, Y., 2017. The financial1336feasibility of producing fish and vegetables through aquaponics. Aquacultural Engineering. 78, 146-1337154.
- Bostock, J., McAndrew, B., Richards, R., Jauncey, K., Telfer, T., Lorenzen, K., Little, D., Ross, L., Handisyde, N.,
 Gatward, I., Corner, R., 2010. Aquaculture: global status and trends. Philosophical Transactions of the
 Royal Society of London B: Biological Sciences. 365, 2897-2912.
- Boyd, C., 1990. Water quality in ponds for aquaculture. Alabama Argicultural Experiment Station, Auburn.
- 1342Boyd, C.E., Chainark, S., 2009. Advances in technology and practice for land-based aquaculture systems:1343Ponds for finfish production. in: Burnel, G., Allen, G. (Eds.), New Technologies in Aquaculture:1344Improving Production Efficiency, Quality and Environmental Management. Woodhead Publishing Ltd.,1345Camebridge, U.K., pp. 984-1009.
- 1346Boyd, C.E., Tucker, C., McNevin, A., Bostick, K., Clay, J., 2007. Indicators of resource use efficiency and1347environmental performance in fish and crustacean aquaculture. Reviews in Fisheries Science. 15, 327-1348360.
- 1349Boyd, C.E., D'Abramo, L.R., Glencross, B.D., Huyben, D.C., Juarez, L.M., Lockwood, G.S., McNevin, A.A.,1350Tacon, A.G.J., Teletchea, F., Tomasso, J.R., Jr., Tucker, C.S., Valenti, W.C., 2020. Achieving

- 1351sustainable aquaculture: Historical and current perspectives and future needs and challenges. Journal1352of the World Aquaculture Society. 51, 578-633.
- 1353Bregnballe, J., 2015. A Guide to Recirculation Aquaculture: An Introduction to the New Environmentally Friendly1354and Highly Productive Closed Fish Farming Systems. A Guide to Recirculation Aquaculture.
- Broch, O.J., Ellingsen, I.H., Forbord, S., Wang, X., Volent, Z., Alver, M.O., Handå, A., Andresen, K., Slagstad,
 D., Reitan, K.I., Olsen, Y., Skjermo, J., 2013. Modelling the cultivation and bioremediation potential of
 the kelp Saccharina latissima in close proximity to an exposed salmon farm in Norway. Aquacult.
 Environ. Interact. 4, 187-206.
- Buschmann, A.H., Camus, C., Infante, J., Neori, A., Israel, Á., Hernández-González, M.C., Pereda, S.V., Gomez Pinchetti, J.L., Golberg, A., Tadmor-Shalev, N., Critchley, A.T., 2017. Seaweed production: overview of
 the global state of exploitation, farming and emerging research activity. European Journal of
 Phycology. 52, 391-406.
- Castine, S.A., Bogard, J.R., Barman, B.K., Karim, M., Mokarrom Hossain, M., Kunda, M., Mahfuzul Haque,
 A.B.M., Phillips, M.J., Thilsted, S.H., 2017. Homestead pond polyculture can improve access to
 nutritious small fish. Food Security. 9, 785-801.
- 1366Cheney, D., Langan, R., Heasman, K., Friedman, B., Davis, J., 2010. Shellfish culture in the open ocean:1367Lessons learned for offshore expansion. Mar. Technol. Soc. J. 44, 55-67.
- 1368Cheng, Y., Wu, X., Li, J., 2018. Chinese mitten Crab Culture: Current Status and Recent Progress Towards1369Sustainable Development, Aquaculture in China: Success Stories and Modern Trends, pp. 197-217.
- 1370 Chopin, T., 2010. Integrated Multi-Trophic Aquaculture. OECD Publishing, pp. 184 205.
- 1371Chopin, T., Tacon, A.G.J., 2020. Importance of Seaweeds and Extractive Species in Global Aquaculture1372Production. Reviews in Fisheries Science & Aquaculture, 1-10.
- 1373 Chopin, T., Cooper, J.A., Reid, G., Cross, S., Moore, C., 2012. Open-water integrated multi-trophic aquaculture:
 1374 environmental biomitigation and economic diversification of fed aquaculture by extractive aquaculture.
 1375 Reviews in Aquaculture. 4, 209-220.
- 1376 Chopin, T., Buschmann, A.H., Halling, C., Troell, M., Kautsky, N., Neori, A., Kraemer, G.P., Zertuche-González,
 1377 J.A., Yarish, C., Neefus, C., 2001. Integrating seaweeds into marine aquaculture systems: a key
 1378 toward sustainability. Journal of Phycology. 37, 975-986.
- 1379 Chu, Y.I., Wang, C.M., Park, J.C., Lader, P.F., 2020. Review of cage and containment tank designs for offshore 1380 fish farming. Aquaculture. 519.
- Clarke, R., Bostock, J., 2017. Regional review on status and trends in aquaculture development in Europe 2015, FAO Fisheries and Aquaculture Circular, Rome, Italy.
- 1383 Cochrane, K., De Young, C., Soto, D., Bahri, T., 2009. Climate change implications for fisheries and aquaculture. FAO Fisheries and aquaculture technical paper. 530, 212.
- Costello, C., Cao, L., Gelcich, S., Cisneros-Mata, M.Á., Free, C.M., Froehlich, H.E., Golden, C.D., Ishimura, G.,
 Maier, J., Macadam-Somer, I., Mangin, T., Melnychuk, M.C., Miyahara, M., de Moor, C.L., Naylor, R.,
 Nøstbakken, L., Ojea, E., O'Reilly, E., Parma, A.M., Plantinga, A.J., Thilsted, S.H., Lubchenco, J., 2020.
 The future of food from the sea. Nature. 588, 95-100.
- Cottrell, R.S., Blanchard, J.L., Halpern, B.S., Metian, M., Froehlich, H.E., 2020. Global adoption of novel
 aquaculture feeds could substantially reduce forage fish demand by 2030. Nature Food. 1, 301-308.
- Cottrell, R.S., Metian, M., Froehlich, H.E., Blanchard, J.L., Jacobson, N.S., McIntyre, P.B., Nash, K.L., Williams,
 D.R., Bouwman, L., Gephart, J.A., Kuempel, C.D., Moran, D.D., Troell, M., Halpern, B.S., 2021. Time
 to rethink trophic levels in aquaculture policy. Reviews in Aquaculture.
- 1394 Cranford, P.J., Reid, G.K., Robinson, S.M.C., 2013. Open water integrated multi-trophic aquaculture:
 1395 Constraints on the effectiveness of mussels as an organic extractive component. Aquacult. Environ.
 1396 Interact. 4, 163-173.
- 1397 Cremer, M., Chappell, J., 2014. New intensive pond aquaculture technology demonstated in China. Global 1398 Aquaculture Advocate. January- February 2014, 60-62.
- Cuvin-Aralar, M.L.A., Lazartigue, A.G., Aralar, E.V., 2009. Cage culture of the Pacific white shrimp Litopenaeus
 vannamei (Boone, 1931) at different stocking densities in a shallow eutrophic lake. Aquaculture
 Research. 40, 181-187.
- 1402Dalsgaard, J., Lund, I., Thorarinsdottir, R., Drengstig, A., Arvonen, K., Pedersen, P.B., 2013. Farming different1403species in RAS in Nordic countries: Current status and future perspectives. Aquacultural Engineering.140453, 2-13.
- 1405Dauda, A.B., 2020. Biofloc technology: a review on the microbial interactions, operational parameters and1406implications to disease and health management of cultured aquatic animals. Reviews in Aquaculture.140712, 1193-1210.

- David, G.S., Carvalho, E.D., Lemos, D., Silveira, A.N., Dall'Aglio-Sobrinho, M., 2015. Ecological carrying
 capacity for intensive tilapia (Oreochromis niloticus) cage aquaculture in a large hydroelectrical
 reservoir in Southeastern Brazil. Aquacultural Engineering. 66, 30-40.
- 1411De Silva, S.S., 2016. Culture based fisheries in Asia are a strategy to augment food security. Food Security. 8,1412585-596.
- 1413 Delmendo, M.N., Gedney, R.H., 1976. Laguna de Bay fish pen aquaculture development Philippines.
 1414 Proceedings of the Annual Meeting World Mariculture Society. 7, 257-265.
- Diana, J.S., Egna, H.S., Chopin, T., Peterson, M.S., Cao, L., Pomeroy, R., Verdegem, M., Slack, W.T., Bondad Reantaso, M.G., Cabello, F., 2013. Responsible aquaculture in 2050: Valuing local conditions and
 human innovations will be key to success. BioScience. 63, 255-262.
- 1418 Donncha, F.O., Grant, J., 2019. Precision Aquaculture. IEEE Internet of Things Magazine. 2, 26-30.
- 1419 Drózdz, D., Malińska, K., Mazurkiewicz, J., Kacprzak, M., Mrowiec, M., Szczypiór, A., Postawa, P., Stachowiak,
 1420 T., 2020. Fish pond sediment from aquaculture production-current practices and the potential for
 1421 nutrient recovery: A Review. International Agrophysics. 34, 33-41.
- 1422Eck, M., Körner, O., Jijakli, M.H., 2019. Nutrient Cycling in Aquaponics Systems. in: Goddek, S., Joyce, A.,1423Kotzen, B., Burnell, G.M. (Eds.), Aquaponics Food Production Systems: Combined Aquaculture and1424Hydroponic Production Technologies for the Future. Springer International Publishing, Cham, pp. 231-1425246.
- 1426Edwards, P., 2015. Aquaculture environment interactions: Past, present and likely future trends. Aquaculture.1427447, 2-14.
- 1428Eggertsen, M., Halling, C., 2021. Knowledge gaps and management recommendations for future paths of1429sustainable seaweed farming in the Western Indian Ocean. Ambio. 50, 60-73.
- EU-Water-Frame-Directive, 2000. Ditective 2000/60/EC of the European Parlement and of the Council of 23
 Octover 2000 establishing a framwork for Community action in the field of water policy, OJ L 327, 22.12.2000, pp. 1-40.
- 1433 EUMOFA, 2020. Recirculating Aquaculture Sstems, European Market Observatory for Fisheries and Aquaculture 1434 Products, Brussels, pp. 45.
- 1435 FAO, 2016. The State of World Fisheries and Aquaculture 2016. Rome.
- 1436 FAO, 2020. The state of World Fisheries and Aquaculture 2020. Sustainability in Action., Rome.
- 1437FAO/FishStatJ, 2021. Fisheries and aquaculture software. FishStatJ Sorftware for Fishery and Aquaculture1438Statistical Time Series. Bibliographic citation [online]. Version released March 2021.
- Ferreira, J.G., Sequeira, A., Hawkins, A.J.S., Newton, A., Nickell, T.D., Pastres, R., Forte, J., Bodoy, A., Bricker,
 S.B., 2009. Analysis of coastal and offshore aquaculture: Application of the FARM model to multiple
 systems and shellfish species (DOI:10.1016/j.aquaculture.2008.12.017). Aquaculture. 292, 129-138.
- 1442Flegel, T.W., 2019. A future vision for disease control in shrimp aquaculture. Journal of the World Aquaculture1443Society. 50, 249-266.
- 1444Flores-Aguilar, R.A., Gutierrez, A., Ellwanger, A., Searcy-Bernal, R., 2007. Development and current status of
abalone aquaculture in Chile. Journal of Shellfish Research. 26, 705-711.
- Freed, S., Kura, Y., Sean, V., Mith, S., Cohen, P., Kim, M., Thay, S., Chhy, S., 2020a. Rice field fisheries: Wild
 aquatic species diversity, food provision services and contribution to inland fisheries. Fisheries
 Research. 229.
- Freed, S., Barman, B., Dubois, M., Flor, R.J., Funge-Smith, S., Gregory, R., Hadi, B.A.R., Halwart, M., Haque,
 M., Jagadish, S.V.K., Joffre, O.M., Karim, M., Kura, Y., McCartney, M., Mondal, M., Nguyen, V.K.,
 Sinclair, F., Stuart, A.M., Tezzo, X., Yadav, S., Cohen, P.J., 2020b. Maintaining Diversity of Integrated
 Rice and Fish Production Confers Adaptability of Food Systems to Global Change. Frontiers in
 Sustainable Food Systems. 4.
- 1454Froehlich, H.E., Runge, C.A., Gentry, R.R., Gaines, S.D., Halpern, B.S., 2018. Comparative terrestrial feed and1455land use of an aquaculture-dominant world. Proceedings of the National Academy of Sciences of the1456United States of America. 115, 5295-5300.
- Garlock, T., Asche, F., Anderson, J., Bjørndal, T., Kumar, G., Lorenzen, K., Ropicki, A., Smith, M.D., Tveterås,
 R., 2020. A Global Blue Revolution: Aquaculture Growth Across Regions, Species, and Countries.
 Reviews in Fisheries Science and Aquaculture. 28, 107-116.
- Gentry, R.R., Froehlich, H.E., Grimm, D., Kareiva, P., Parke, M., Rust, M., Gaines, S.D., Halpern, B.S., 2017.
 Mapping the global potential for marine aquaculture. Nature Ecology and Evolution. 1, 1317-1324.
- 1462Gephart, J.A., Troell, M., Henriksson, P.J.G., Beveridge, M.C.M., Verdegem, M., Metian, M., Mateos, L.D.,1463Deutsch, L., 2016. The `seafood gap' in the food-water nexus literature-issues surrounding freshwater

- 1464use in seafood production chains. Advances in Water Resources on-line, DOI:146510.1016/j.advwatres.2017.1003.1025.
- Gephart, J.A., Golden, C.D., Asche, F., Belton, B., Brugere, C., Froehlich, H.E., Fry, J.P., Halpern, B.S., Hicks,
 C.C., Jones, R.C., Klinger, D.H., Little, D.C., McCauley, D.J., Thilsted, S.H., Troell, M., Allison, E.H.,
 2020. Scenarios for Global Aquaculture and Its Role in Human Nutrition. Reviews in Fisheries Science
 and Aquaculture. 29, 122-138.
- 1470Ghamkhar, R., Boxman, S.E., Main, K.L., Zhang, Q., Trotz, M.A., Hicks, A., 2021. Life cycle assessment of1471aquaculture systems: Does burden shifting occur with an increase in production intensity? Aquacultural1472Engineering. 92.
- 1473Goddek, S., Espinal, C.A., Delaide, B., Jijakli, M.H., Schmautz, Z., Wuertz, S., Keesman, K.J., 2016. Navigating1474towards Decoupled Aquaponic Systems: A System Dynamics Design Approach. Water. 8, 303.
- 1475 Green, B.S., Gardner, C., Hochmuth, J.D., Linnane, A., 2014. Environmental effects on fished lobsters and 1476 crabs. Reviews in Fish Biology and Fisheries. 24, 613-638.
- Gui, J., Tang, Q., Li, Z., Liu, J., De Silva, S.S., 2018. Aquaculture in China : success stories and modern trends.
 John Wiley & Sons Ltd, [Hoboken, NJ].
- Guillemin, M.L., Faugeron, S., Destombe, C., Viard, F., Correa, J.A., Valero, M., 2008. Genetic variation in wild and cultivated populations of the haploid-diploid red alga Gracilaria chilensis: How farming practices favor asexual reproduction and heterozygosity. Evolution. 62, 1500-1519.
- 1482 Guo, X., 2021. Genetics in shellfish culture. in: Shumway, S.E. (Ed.), Molluscan Shellfish Aquaculture : A 1483 Practical Guide. 5m Publishing, Portland, pp. 393-414.
- Hafting, J.T., Craigie, J.S., Stengel, D.B., Loureiro, R.R., Buschmann, A.H., Yarish, C., Edwards, M.D., Critchley,
 A.T., 2015. Prospects and challenges for industrial production of seaweed bioactives. Journal of
 Phycology. 51, 821-837.
- 1487Halim, D., Juanri, 2016. Indonesians aquaculture industry key sectors for future growth,1488http://www.jpsosconsulting.com/asean-agriculture.
- 1489Hallegraeff, G.M., 2010. Ocean climate change, phytoplankton community responses, and harmful algal1490blooms: A formidable predictive challenge. Journal of Phycology. 46, 220-235.
- 1491Hassan, S.G., Hasan, M., Li, D., 2016. Information fusion in aquaculture: A state-of the art review. Frontiers of1492Agricultural Science and Engineering. 3, 206-221.
- Henriksson, P.J.G., Belton, B., Murshed-E-Jahan, K., Rico, A., 2018. Measuring the potential for sustainable
 intensification of aquaculture in Bangladesh using life cycle assessment. Proceedings of the National
 Academy of Sciences of the United States of America. 115, 2958-2963.
- Henriksson, P.J.G., Dickson, M., Allah, A.N., Al-Kenawy, D., Phillips, M., 2017. Benchmarking the environmental performance of best management practice and genetic improvements in Egyptian aquaculture using life cycle assessment. Aquaculture. 468, 53-59.
- 1499 Hickling, C.F., 1962. Fish culture. [s.n.], London.
- Hilborn, R., Banobi, J., Hall, S.J., Pucylowski, T., Walsworth, T.E., 2018. The environmental cost of animal source foods. Frontiers in Ecology and the Environment. 16, 329-335.
- Hu, Z., Lee, J.W., Chandran, K., Kim, S., Khanal, S.K., 2012. Nitrous oxide (N 2O) emission from aquaculture:
 A review. Environmental Science and Technology. 46, 6470-6480.
- Hua, K., Cobcroft, J.M., Cole, A., Condon, K., Jerry, D.R., Mangott, A., Praeger, C., Vucko, M.J., Zeng, C.,
 Zenger, K., Strugnell, J.M., 2019. The Future of Aquatic Protein: Implications for Protein Sources in
 Aquaculture Diets. One Earth. 1, 316-329.
- Huang, A., 2019. RAS in China. in: Dalsgaard, J. (Ed.), 5th NordicRAS workshop on recirculating aquaculture
 systems, Book of Abstracts. DTU Aqua Report No. 350-2019. National Institute of Aqutic Resources,
 Technical University of Denmark, Berlin, Germany, pp. 50.
- 1510 Huet, M., 1986. Textbook of fish culture : breeding and cultivation of fish. Fishing News Books, Farnham.
- 1511Hughes, A.D., Black, K.D., 2016. Going beyond the search for solutions: Understanding trade-offs in European1512integrated multi-trophic aquaculture development. Aquacult. Environ. Interact. 8, 191-199.
- 1513Hurd, C.L., Lobban, C.S., Bischof, K., Harrison, P.J., 2014. Seaweed mariculture (Chapter 10), In Seaweed1514Ecology and Physiology. Cambridge University Press, Cambridge, pp. 413-439.
- Hurtado, A.Q., Neish, I.C., Critchley, A.T., 2019. Phyconomy: the extensive cultivation of seaweeds, their
 sustainability and economic value, with particular reference to important lessons to be learned and
 transferred from the practice of eucheumatoid farming. Phycologia. 58, 472-483.

- Husen, M.A., Yadav, C.N.R., Shreshtha, M., Bista, J.D., 2012. Growth and production of planktivorous fish
 species in cages stocked as monoculture and polyculture at Khapuadi in Phewa Lake, Nepal. Asian
 Fish. Sci. 25, 218-231.
- 1521 Hwang, E.K., Park, C.S., 2020. Seaweed cultivation and utilization of Korea. Algae. 35, 107-121.
- 1522 Infante-Villamil, S., Huerlimann, R., Jerry, D.R., 2020. Microbiome diversity and dysbiosis in aquaculture. 1523 Reviews in Aquaculture.
- Itano, T., Inagaki, T., Nakamura, C., Hashimoto, R., Negoro, N., Hyodo, J., Honda, S., 2019. Water circulation induced by mechanical aerators in a rectangular vessel for shrimp aquaculture. Aquacultural Engineering. 85, 106-113.
- Jansen, H.M., Van Den Burg, S., Bolman, B., Jak, R.G., Kamermans, P., Poelman, M., Stuiver, M., 2016. The feasibility of offshore aquaculture and its potential for multi-use in the North Sea. Aquaculture International. 24, 735-756.
- Jayanthi, M., Balasubramaniam, A.A.K., Suryaprakash, S., Veerapandian, N., Ravisankar, T., Vijayan, K.K.,
 2021. Assessment of standard aeration efficiency of different aerators and its relation to the overall
 economics in shrimp culture. Aquacultural Engineering. 92.
- 1533Joffre, O.M., Verdegem, M.C.J., 2019. Feeding both pond and fish: a pathway to ecological intensification of1534aquaculture systems. Infofish International (www.infofish.org), 60-63.
- Jokumsen, A., Svendsen, L.M., 2010. Farming of freshwater rainbow trout in Denmark, DTU Aqua Report no.
 219-2010. DTU Aqua Technical University Denmark.
- Kabir, K.A., Schrama, J.W., Verreth, J.A.J., Phillips, M.J., Verdegem, M.C.J., 2019. Effect of dietary protein to
 energy ratio on performance of Nile tilapia and food web enhancement in semi-intensive pond
 aquaculture. Aquaculture. 499, 235-242.
- Kabir, K.A., Verdegem, M.C.J., Verreth, J.A.J., Phillips, M.J., Schrama, J.W., 2020. Dietary non-starch
 polysaccharides influenced natural food web and fish production in semi-intensive pond culture of Nile
 tilapia. Aquaculture. 528.
- 1543 Kawahigashi, D., 2018. New paradigm for controlling EMS/APHNS in intensive p. Vannamei boone 1931 culture 1544 ponds. Asian Fisheries Science. 31S, 182-193.
- 1545 Kinyage, J.P.H., Pedersen, P.B., Pedersen, L.F., 2019. Effects of abrupt salinity increase on nitrification 1546 processes in a freshwater moving bed biofilter. Aquacultural Engineering. 84, 91-98.
- 1547 Klinger, D., Naylor, R., 2012. Searching for solutions in aquaculture: Charting a sustainable course, Annual 1548 Review of Environment and Resources, pp. 247-276.
- 1549 Klinger, D.H., Levin, S.A., Watson, J.R., 2017. The growth of finfish in global openocean aquaculture under 1550 climate change. Proceedings of the Royal Society B: Biological Sciences. 284.
- 1551Kumar, A., Moulick, S., Mal, B.C., 2013. Selection of aerators for intensive aquacultural pond. Aquacultural1552Engineering. 56, 71-78.
- 1553 Kumar, V., Roy, S., Meena, D.K., Sarkar, U.K., 2016. Application of probiotics in shrimp aquaculture:
 1554 importance, mechanisms of action, and methods of administration. Reviews in Fisheries Science & Aquaculture. 24, 342-368.
- 1556 Latt, U.W., 2019. Biosecurity application in shrimp hatchery: beneficial ar a cost factor. Presentation made at 1557 *OIE Global Conference on aquatic Animal Helath,* Santiago, Chile, 2-4 April 2019.
- 1558Lebel, L., Navy, H., Jutagate, T., Akester, M.J., Sturm, L., Lebel, P., Lebel, B., 2020. Innovation, Practice, and1559Adaptation to Climate in the Aquaculture Sector. Reviews in Fisheries Science and Aquaculture.
- Lepine, C., Christianson, L., Soucek, D., McIsaac, G., Summerfelt, S., 2021. Metal leaching and toxicity of denitrifying woodchip bioreactor outflow—Potential reuse application. Aquacultural Engineering. 93, 102129.
- Li, R.H., Liu, S.M., Zhang, J., Jiang, Z.J., Fang, J.G., 2016. Sources and export of nutrients associated with integrated multi-trophic aquaculture in Sanggou Bay, China. Aquacult. Environ. Interact. 8, 285-309.
- Lindholm-Lehto, P., Pulkkinen, J., Kiuru, T., Koskela, J., Vielma, J., 2020. Water quality in recirculating
 aquaculture system using woodchip denitrification and slow sand filtration. Environmental Science and
 Pollution Research. 27, 17314-17328.
- Little, D.C., Newton, R.W., Beveridge, M.C.M., 2016. Aquaculture: A rapidly growing and significant source of sustainable food? Status, transitions and potential. Proceedings of the Nutrition Society. 75, 274-286.
- Lombardi, J.V., de Almeida Marques, H.L., Pereira, R.T.L., Barreto, O.J.S., de Paula, E.J., 2006. Cage
 polyculture of the Pacific white shrimp Litopenaeus vannamei and the Philippines seaweed
 Kappaphycus alvarezii. Aquaculture. 258, 412-415.

- 1573 López-Vivas, J.M., Riosmena-Rodríguez, R., de la Llave, A.A.J.-G., Pacheco-Ruíz, I., Yarish, C., 2015. Growth
 1574 and reproductive responses of the conchocelis phase of Pyropia hollenbergii (Bangiales, Rhodophyta)
 1575 to light and temperature. Journal of Applied Phycology. 27, 1561-1570.
- Luo, Z., Hu, S., Chen, D., 2018. The trends of aquacultural nitrogen budget and its environmental implications in China. Scientific Reports. 8.
- 1578 Maas, R.M., Verdegem, M.C., Dersjant-Li, Y., Schrama, J.W., 2018. The effect of phytase, xylanase and their 1579 combination on growth performance and nutrient utilization in Nile tilapia. Aquaculture. 487, 7-14.
- 1580Maas, R.M., Verdegem, M.C.J., Wiegertjes, G.F., Schrama, J.W., 2020. Carbohydrate utilisation by tilapia: a1581meta-analytical approach. Reviews in Aquaculture.
- Maas, R.M., Verdegem, M.C.J., Debnath, S., Marchal, L., Schrama, J.W., 2021. Effect of enzymes (phytase and xylanase), probiotics (B. amyloliquefaciens) and their combination on growth performance and nutrient utilisation in Nile tilapia. Aquaculture. 533, 736226.
- 1585 MacLeod, M.J., Hasan, M.R., Robb, D.H.F., Mamun-Ur-Rashid, M., 2020. Quantifying greenhouse gas emissions 1586 from global aquaculture. Scientific Reports. 10, 11679.
- 1587Mahmood, T., Fang, J., Jiang, Z., Zhang, J., 2016a. Seasonal nutrient chemistry in an integrated multi-trophic1588aquaculture region: case study of Sanggou Bay from North China. Chem. Ecol. 32, 149-168.
- Mahmood, T., Fang, J.G., Jiang, Z.J., Zhang, J., 2016b. Carbon and nitrogen flow, and trophic relationships, among the cultured species in an integrated multi-trophic aquaculture (IMTA) bay. Aquacult. Environ. Interact. 8, 207-219.
- Maiolo, S., Parisi, G., Biondi, N., Lunelli, F., Tibaldi, E., Pastres, R., 2020. Fishmeal partial substitution within
 aquafeed formulations: life cycle assessment of four alternative protein sources. International Journal
 of Life Cycle Assessment. 25, 1455-1471.
- Martínez-Porchas, M., Martínez-Córdova, L.R., Porchas-Cornejo, M.A., López-Elías, J.A., 2010. Shrimp
 polyculture: A potentially profitable, sustainable, but uncommon aquacultural practice. Reviews in
 Aquaculture. 2, 73-85.
- Martins, C., Eding, E.H., Verdegem, M.C., Heinsbroek, L.T., Schneider, O., Blancheton, J.-P., d'Orbcastel, E.R.,
 Verreth, J., 2010. New developments in recirculating aquaculture systems in Europe: A perspective on
 environmental sustainability. Aquacultural engineering. 43, 83-93.
- 1601 Masso-Silva, J.A., Diamond, G., 2014. Antimicrobial Peptides from Fish. Pharmaceuticals. 7, 265-310.
- 1602 Matassa, S., Boon, N., Pikaar, I., Verstraete, W., 2016. Microbial protein: future sustainable food supply route 1603 with low environmental footprint. Microb. Biotechnol. 9, 568-575.
- Matsuyama, Y., Shumway, S., 2009. Impacts of harmful algal blooms on shellfisheries aquaculture, New
 Technologies in Aquaculture. Elsevier, pp. 580-609.
- Maucieri, C., Nicoletto, C., Os, E.v., Anseeuw, D., Havermaet, R.V., Junge, R., 2019. Hydroponic Technologies.
 in: Goddek, S., Joyce, A., Kotzen, B., Burnell, G.M. (Eds.), Aquaponics Food Production Systems:
 Combined Aquaculture and Hydroponic Production Technologies for the Future. Springer International
 Publishing, Cham, pp. 77-110.
- Merican, Z., 2021. Shrimp broodstock and hatchery in 2020: making waves with fast growth lines., AQUA
 Cylture Asia Pacific. , pp. 6-11.
 <u>https://issuu.com/aguacultureasiapacific/docs/ag21153_aap_janfeb21121_fa_sml</u>.
- 1613 Midilli, A., Kucuk, H., Dincer, I., 2012. Environmental and sustainability aspects of a recirculating aquaculture 1614 system. Environmental Progress & Sustainable Energy. 31, 604-611.
- Millard, R.S., Ellis, R.P., Bateman, K.S., Bickley, L.K., Tyler, C.R., van Aerle, R., Santos, E.M., 2020. How do
 abiotic environmental conditions influence shrimp susceptibility to disease? A critical analysis focussed
 on White Spot Disease. Journal of Invertebrate Pathology.
- Molina, C., Espinoza, M., 2018. Rising use of automatic feeders in shrimp ponds poses new feed requirements.,
 Global Aquaculture Advocate, 20 August 2018,
 <u>https://www.aquaculturealliance.org/advocate/automatic-feeders-shrimp-ponds-new-feed-</u>
 <u>requirements/?headlessPrint=AAAAAPIA9c8r7gs82oWZB</u>.
- 1622Monsees, H., Kloas, W., Wuertz, S., 2017. Decoupled systems on trial: Eliminating bottlenecks to improve1623aquaponic processes. PLoS ONE. 12.
- 1624NACA/FAO, 2000. Aquaculture development beyond 2000: the Bangkok Declaration and Strategy, Conference1625on Aquaculture in the Third Millenium. NACA, Bangkok and FAO, Rome. 27 pp., Bangkok, Thailand.
- Naylor, R.L., Goldburg, R.J., Primavera, J.H., Kautsky, N., Beveridge, M.C.M., Clay, J., Folke, C., Lubchenco, J.,
 Mooney, H., Troell, M., 2000. Effect of aquaculture on world fish supplies. Nature. 405, 1017-1024.

- Naylor, R.L., Hardy, R.W., Buschmann, A.H., Bush, S.R., Cao, L., Klinger, D.H., Little, D.C., Lubchenco, J.,
 Shumway, S.E., Troell, M., 2021. A 20-year retrospective review of global aquaculture. Nature. 591,
 551-563.
- Naylor, R.L., Hardy, R.W., Bureau, D.P., Chiu, A., Elliott, M., Farrell, A.P., Forster, I., Gatlin, D.M., Goldburg,
 R.J., Hua, K., Nichols, P.D., 2009. Feeding aquaculture in an era of finite resources. Proceedings of the
 National Academy of Sciences of the United States of America. 106, 15103-15110.
- Neori, A., Troell, M., Chopin, T., Yarish, C., Critchley, A., Buschmann, A.H., 2007. The need for a balanced
 ecosystem approach to blue revolution aquaculture. Environment: Science and Policy for Sustainable
 Development. 49, 36-43.
- 1637 Nhung, T.T., Le Vo, P., Van Nghi, V., Bang, H.Q., 2019. Salt intrusion adaptation measures for sustainable
 agricultural development under climate change effects: A case of Ca Mau Peninsula, Vietnam. Climate
 Risk Management. 23, 88-100.
- 1640 Nhut, N., Hao, N.V., Bosma, R.H., Verreth, J.A.V., Eding, E.H., Verdegem, M.C.J., 2019. Options to reuse
 1641 sludge from striped catfish (Pangasianodon hypophthalmus, Sauvage, 1878) ponds and recirculating
 1642 systems. Aquacultural Engineering. 87.
- 1643 Ninawe, A.S., Selvin, J., 2009. Probiotics in shrimp aquaculture: avenues and challenges. Crit. Rev. Microbiol.
 1644 35, 43-66.
- 1645 Oyinlola, M.A., Reygondeau, G., Wabnitz, C.C.C., Troell, M., Cheung, W.W.L., 2018. Global estimation of areas 1646 with suitable environmental conditions for mariculture species. PLoS ONE. 13.
- 1647Paclibare, J.O., Verdegem, M.C., van Muiswinkel, W., Huisman, B., 1998. The potential for crop rotation in1648controlling diseases in shrimp culture. Naga, the ICLARM Quarterly. 21, 22-24.
- Palm, H.W., Knaus, U., Appelbaum, S., Goddek, S., Strauch, S.M., Vermeulen, T., Haïssam Jijakli, M., Kotzen,
 B., 2018. Towards commercial aquaponics: a review of systems, designs, scales and nomenclature.
 Aquaculture International. 26, 813-842.
- Parra, L., Lloret, G., Lloret, J., Rodilla, M., 2018. Physical Sensors for Precision Aquaculture: A Review. IEEE
 Sens. J. 18, 3915-3923.
- 1654Paudel, S.R., 2020. Nitrogen transformation in engineered aquaponics with water celery (Oenanthe javanica)1655and koi carp (Cyprinus carpio): Effects of plant to fish biomass ratio. Aquaculture. 520, 734971.
- Pelletier, N., Klinger, D.H., Sims, N.A., Yoshioka, J.R., Kittinger, J.N., 2018. Nutritional Attributes,
 Substitutability, Scalability, and Environmental Intensity of an Illustrative Subset of Current and Future
 Protein Sources for Aquaculture Feeds: Joint Consideration of Potential Synergies and Trade-offs.
 Environmental Science and Technology. 52, 5532-5544.
- Pinho, S.M., Molinari, D., de Mello, G.L., Fitzsimmons, K.M., Coelho Emerenciano, M.G., 2017. Effluent from a biofloc technology (BFT) tilapia culture on the aquaponics production of different lettuce varieties. Ecological Engineering. 103, 146-153.
- 1663Rambags, F., Tanner, C.C., Stott, R., Schipper, L.A., 2019. Bacteria and virus removal in denitrifying1664bioreactors: Effects of media type and age. Ecological Engineering. 138, 46-53.
- 1665Raworth, K., 2017. Doughnut economics: seven ways to think like a 21st-century economist. Chelsea Green1666Publishing.
- 1667 Reid, G.K., Forster, I., Cross, S., Pace, S., Balfry, S., Dumas, A., 2017. Growth and diet digestibility of cultured
 1668 sablefish Anoplopoma fimbria : Implications for nutrient waste production and Integrated Multi-Trophic
 1669 Aquaculture. Aquaculture. 470, 223-229.
- Reid, G.K., Gurney-Smith, H.J., Marcogliese, D.J., Knowler, D., Benfey, T., Garber, A.F., Forster, I., Chopin, T.,
 Brewer-Dalton, K., Moccia, R.D., Flaherty, M., Smith, C.T., De Silva, S., 2019. Climate change and
 aquaculture: Considering biological response and resources. Aquacult. Environ. Interact. 11, 569-602.
- 1673 Reverter, M., Tapissier-Bontemps, N., Sarter, S., Sasal, P., Caruso, D., 2021. Moving towards more sustainable
 1674 aquaculture practices: a meta-analysis on the potential of plant-enriched diets to improve fish growth,
 1675 immunity and disease resistance. Reviews in Aquaculture. 13, 537-555.
- Rowley, A.F., Cross, M.E., Culloty, S.C., Lynch, S.A., Mackenzie, C.L., Morgan, E., O'Riordan, R.M., Robins, P.E.,
 Smith, A.L., Thrupp, T.J., Vogan, C.L., Wootton, E.C., Malham, S.K., 2014. The potential impact of
 climate change on the infectious diseases of commercially important shellfish populations in the Irish
 Sea A review. ICES Journal of Marine Science. 71, 741-759.
- Salazar, C., Jaime, M., Figueroa, Y., Fuentes, R., 2018. Innovation in small-scale aquaculture in Chile.
 Aquaculture Economics and Management. 22, 151-167.
- Samuel-Fitwi, B., Nagel, F., Meyer, S., Schroeder, J., Schulz, C., 2013. Comparative life cycle assessment (LCA)
 of raising rainbow trout (Oncorhynchus mykiss) in different production systems. Aquacultural
 engineering. 54, 85-92.

- 1685 Santelices, B., 1999. A conceptual framework for marine agronomy. Hydrobiologia. 398-399, 15-23.
- Sarkis, S., Karney, R., Creswell, R.L., 2021. Design and construction considerations for a molluscan hatchery.
 in: Shumway, S.E. (Ed.), Molluscan Shellfish Aquaculture: A practical guide. 5 M Publications, London.
 In press.
- 1689Schneider, O., Sereti, V., Eding, E., Verreth, J., 2005. Analysis of nutrient flows in integrated intensive1690aquaculture systems. Aquacultural engineering. 32, 379-401.
- 1691 Shinn, A.P., Pratoomyot, J., Griffiths, D., Trong, T.Q., Vu, N.T., Jiravanichpaisal, P., Briggs, M., 2018. Asian 1692 shrimp production and the economic costs of disease. Asian Fisheries Science. 31, 29-58.
- 1693 Shumway, S., in press. Molluscan Shellfish Aquaculture: A practical guide. 5MBooks.
- 1694Shumway, S.E., 1990. A Review of the Effects of Algal Blooms on Shellfish and Aquaculture. Journal of the1695World Aquaculture Society. 21, 65-104.
- 1696 Shumway, S.E., 2011. Shellfish aquaculture and the environment. John Wiley & Sons.
- Silva, C., Ferreira, J.G., Bricker, S.B., DelValls, T.A., Martín-Díaz, M.L., Yáñez, E., 2011. Site selection for
 shellfish aquaculture by means of GIS and farm-scale models, with an emphasis on data-poor
 environments. Aquaculture. 318, 444-457.
- Song, X., Liu, Y., Pettersen, J.B., Brandão, M., Ma, X., Røberg, S., Frostell, B., 2019. Life cycle assessment of recirculating aquaculture systems: A case of Atlantic salmon farming in China. J. Ind. Ecol. 23, 1077-1086.
- Stentiford, G.D., Neil, D.M., Peeler, E.J., Shields, J.D., Small, H.J., Flegel, T.W., Vlak, J.M., Jones, B., Morado,
 F., Moss, S., Lotz, J., Bartholomay, L., Behringer, D.C., Hauton, C., Lightner, D.V., 2012. Disease will
 limit future food supply from the global crustacean fishery and aquaculture sectors. Journal of
 Invertebrate Pathology. 110, 141-157.
- 1707 Stokstad, E., 2020. Tomorrow's catch. Science 370, 902-905.
- Subasinghe, R., Arthur, J., Bartley, D., De Silva, S., Halwart, M., Hishamunda, N., Mohan, C., Sorgeloos, P.,
 2013. Proceedings of the Global Conference on Aquaculture 2010. Farming the waters for people and
 food, Proceedings of the Global Conference on Aquaculture 2010. Farming the waters for people and
 food. FAO/NACA.
- 1712Tacon, A.G.J., 2020. Trends in Global Aquaculture and Aquafeed Production: 2000–2017. Reviews in Fisheries1713Science and Aquaculture. 28, 43-56.
- Thilsted, S.H., Thorne-Lyman, A., Webb, P., Bogard, J.R., Subasinghe, R., Phillips, M.J., Allison, E.H., 2016.
 Sustaining healthy diets: The role of capture fisheries and aquaculture for improving nutrition in the post-2015 era. Food Policy. 61, 126-131.
- 1717Tinh, T.H., Koppenol, T., Hai, T.N., Verreth, J.A.J., Verdegem, M.C.J., 2021. Effects of carbohydrate sources on1718a biofloc nursery system for whiteleg shrimp (Litopenaeus vannamei). Aquaculture. 531.
- Trainer, V.L., Moore, S.K., Hallegraeff, G., Kudela, R.M., Clement, A., Mardones, J.I., Cochlan, W.P., 2020.
 Pelagic harmful algal blooms and climate change: Lessons from nature's experiments with extremes.
 Harmful Algae. 91, 101591.
- 1722 Troell, M., Metian, M., Beveridge, M., Verdegem, M., Deutsch, L., 2014a. Comment on 'Water footprint of 1723 marine protein consumption-aquaculture's link to agriculture'. Environmental Research Letters. 9.
- Troell, M., Joyce, A., Chopin, T., Neori, A., Buschmann, A.H., Fang, J.-G., 2009. Ecological engineering in aquaculture—Potential for integrated multi-trophic aquaculture (IMTA) in marine offshore systems.
 Aquaculture. 297, 1-9.
- Troell, M., Halling, C., Neori, A., Chopin, T., Buschmann, A., Kautsky, N., Yarish, C., 2003. Integrated
 mariculture: asking the right questions. Aquaculture. 226, 69-90.
- Troell, M., Naylor, R.L., Metian, M., Beveridge, M., Tyedmers, P.H., Folke, C., Arrow, K.J., Barrett, S., Crépin,
 A.S., Ehrlich, P.R., Gren, A., Kautsky, N., Levin, S.A., Nyborg, K., Österblom, H., Polasky, S., Scheffer,
 M., Walker, B.H., Xepapadeas, T., De Zeeuw, A., 2014b. Does aquaculture add resilience to the global
 food system? Proceedings of the National Academy of Sciences of the United States of America. 111,
 13257-13263.
- 1734 Tucker, C., Hargreaves, J., 2012. Ponds. in: Tidwell, J.H. (Ed.), Aquaculture Production Systems, pp. 191-244.
- 1735 Uccello, E., Kauffmann, D., Calo, M., Streissel, M., 2017. Nutrition-sensitive agriculture and food systems in 1736 practice: options for intervention. FAO.
- Usandizaga, S., Buschmann, A.H., Camus, C., Kappes, J.L., Arnaud-Haond, S., Mauger, S., Valero, M.,
 Guillemin, M.L., 2020. Better off alone? Compared performance of monoclonal and polyclonal stands of
 a cultivated red alga growth. Evol. Appl. 13, 905-917.

- Valero, M., Guillemin, M.L., Destombe, C., Jacquemin, B., Gachon, C.M.M., Badis, Y., Buschmann, A.H., Camus,
 C., Faugeron, S., 2017. Perspectives on domestication research for sustainable seaweed aquaculture.
 Perspectives in Phycology. 4, 33-46.
- 1743 Van Rijn, J., 2013. Waste treatment in recirculating aquaculture systems. Aquacultural Engineering. 53, 49-56.
- 1744 Van Zanten, H.H.E., Van Ittersum, M.K., De Boer, I.J.M., 2019. The role of farm animals in a circular food
 1745 system. Global Food Security. 21, 18-22.
- 1746 Verdegem, M.C.J., Bosma, R.H., 2009. Water withdrawal for brackish and inland aquaculture, and options to 1747 produce more fish in ponds with present water use. Water Policy. 11, 52-68.
- Verdegem, M.C.J., Bosma, R.H., Verreth, J.A.J., 2006. Reducing water use for animal production through aquaculture. International Journal of Water Resources Development. 22, 101-113.
- von Ahnen, M., Pedersen, P.B., Dalsgaard, J., 2018. Performance of full-scale woodchip bioreactors treating
 effluents from commercial RAS. Aquacultural Engineering. 83, 130-137.
- Wang, Q., Ding, H., Tao, Z., Ma, D., 2018. Crayfish (Procambarus clarkii) Cultivation in China: A Decade of
 Unprecedented Development, Aquaculture in China: Success Stories and Modern Trends, pp. 363-377.
- Wang, X., LM, O., KI, R., Y, O., 2012. Discharge of nutrient wastes from salmon farms: environmental effects, and potential for integrated multi-trophic aquaculture. Aquacult. Environ. Interact. 2, 267-283.
- Watson, D., Shumway, S., Whitlatch, R., 2009. Biofouling and the shellfish industry, Shellfish Safety and quality. Elsevier, pp. 317-337.
- Weitzman, J., 2019. Applying the ecosystem services concept to aquaculture: A review of approaches, definitions, and uses. Ecosystem Services. 35, 194-206.
- Wells, M.L., Potin, P., Craigie, J.S., Raven, J.A., Merchant, S.S., Helliwell, K.E., Smith, A.G., Camire, M.E., Brawley, S.H., 2017. Algae as nutritional and functional food sources: revisiting our understanding. Journal of Applied Phycology. 29, 949-982.
- 1763 Wilfart, A., Prudhomme, J., Blancheton, J.P., Aubin, J., 2013. LCA and emergy accounting of aquaculture 1764 systems: Towards ecological intensification. J. Environ. Manage. 121, 96-109.
- Wongkiew, S., Hu, Z., Chandran, K., Lee, J.W., Khanal, S.K., 2017. Nitrogen transformations in aquaponic
 systems: A review. Aquacultural Engineering. 76, 9-19.
- Xiao, R., Wei, Y., An, D., Li, D., Ta, X., Wu, Y., Ren, Q., 2019. A review on the research status and development trend of equipment in water treatment processes of recirculating aquaculture systems. Reviews in Aquaculture. 11, 863-895.
- Xiao, X., Agusti, S., Lin, F., Li, K., Pan, Y., Yu, Y., Zheng, Y., Wu, J., Duarte, C.M., 2017. Nutrient removal from Chinese coastal waters by large-scale seaweed aquaculture. Scientific Reports. 7.
- Xiao, X., Agustí, S., Yu, Y., Huang, Y., Chen, W., Hu, J., Li, C., Li, K., Wei, F., Lu, Y., Xu, C., Chen, Z., Liu, S.,
 Zeng, J., Wu, J., Duarte, C.M., 2021. Seaweed farms provide refugia from ocean acidification. Science of the Total Environment. 776.
- Yang, P., Zhang, Y., Yang, H., Guo, Q., Lai, D.Y.F., Zhao, G., Li, L., Tong, C., 2020. Ebullition was a major
 pathway of methane emissions from the aquaculture ponds in southeast China. Water Research. 184.
- 1777Yang, Y., Chai, Z., Wang, Q., Chen, W., He, Z., Jiang, S., 2015. Cultivation of seaweed Gracilaria in Chinese1778coastal waters and its contribution to environmental improvements. Algal Research. 9, 236-244.
- Yi, Y., Fitzsimmons, K., 2004. Survey of tilapia-shrimp polyculture in Thailand. in: Harris, R., Courter, I., Egna,
 H. (Eds.), Twenty-First Annual Technical Report. Oregon State University, Corvalis, Oregon, USA.
- 1781 Zhang, Y., Bleeker, A., Liu, J., 2015. Nutrient discharge from China's aquaculture industry and associated
 1782 environmental impacts. Environmental Research Letters. 10.
- 1783 Zhou, C., Xu, D., Lin, K., Sun, C., Yang, X., 2018. Intelligent feeding control methods in aquaculture with an 1784 emphasis on fish: a review. Reviews in Aquaculture. 10, 975-993.